Meeting Educational Needs with “Course” Correction

Remodelled M.Sc. Biotechnology Curriculum
Meeting Educational Needs with "Course" Correction
Remodelled M.Sc. Biotechnology Curriculum
Meeting Educational Needs
with “Course” Correction
Remodelled M.Sc. Biotechnology Curriculum

May, 2017

Copyright © Deptt. of Biotechnology
Ministry of Science & Technology
Government of India

Compiled and Coordinated
Ms. Shreya Malik, DM, BCIL.

Edited
Dr. Suman Govil, Adviser, DBT
Dr. Purnima Sharma, MD, BCIL

Assisted
Mr. Naman Khare, PE, BCIL

Published
Biotech Consortium India Limited
http://bcil.nic.in/

Designed
Ms. Shweta
+91 9930774161

Printed
M/s. Mittal Enterprises, Delhi 110006
+91 9811340726
MESSAGE

Dr. Harsh Vardhan
Minister for Science & Technology and Earth Sciences

Department of Biotechnology initiated Integrated Human Resource Development Programme way back in 1985-86 to cater to the requirement of quality manpower for R&D, teaching and manufacturing activities. I am very proud that India is one of the first countries in the world to initiate postgraduate teaching programme in Biotechnology. M.Sc./M.Tech. programme was initiated in 5 universities and has been expanded to over 70 universities/IITs in the country to cover general, medical, agricultural, veterinary, environmental, industrial, marine, food, pharmaceutical biotech.

Students for these programmes are selected on the basis of an All India entrance test and all selected students are paid studentships. I am very happy to know that the Department has initiated major curriculum revision exercise for specialisations offered under DBT supported teaching programme. The exercise has been coordinated by Biotech Consortium India Limited. The Department invited feedback from researchers, academic community, biotech industries and past as well as present students. Feedback has been considered by the Expert groups and areas with recent developments have been included and identified gap areas which need inclusion and updation have been taken care of. I compliment the Department for taking up this major exercise for the benefit of student community and congratulate the group for bringing out this publication.

(Dr. Harsh Vardhan)
Andy Hargreaves, a renowned educational expert, has once remarked, “Capacity building originally meant helping people to help themselves. Now it means required trainee to deliver imposed policies.” In the Indian context, the Integrated Human Resource Development Programme of the Department of Biotechnology is a flagship and dynamic programme that has done exceedingly well to meet the requirements of capacity building. The central idea should be to take enough care in the selection of quality students and provide hands-on practical training to students.

I am extremely happy to note that the Department is revising the curriculum for various PG programmes in Biotechnology at regular intervals to incorporate the latest developments in the field. While doing so, I am told that Biotech Consortium India Limited has obtained necessary feedback from different stakeholders, including researchers, academia, industries, and students regarding the proposed changes in the curriculum. Feedback was analyzed and considered by the Expert Groups with curricula followed by the best international universities. I am assured that the proposed curricula have incorporated papers on research methodology, scientific communication, prevailing regulations in the country, etc.

I am confident that this curriculum revision exercise would be very beneficial for faculty and students of not only DBT-supported programmes but also other universities involved in biotechnology teaching. I compliment the Department for undertaking this valuable exercise.
MESSAGE

Integrated Human Resource Development Programme in biotechnology is a unique, innovative initiative taken by Department of Biotechnology way back in 1985-86. Human Resource Development programmes of the Department are highly dynamic and have evolved continuously based on need, regional aspirations and feedback from different stakeholders.

Emphasis is laid on selection of institutions based on existing expertise, infrastructure, nearby institutions engaged in research in relevant areas and students are provided hands on practical training. These programmes are continuously mentored and monitored by Advisory Committee, Expert Task Force and Course Coordinators meeting. An attempt is made to conduct curriculum revision exercise at frequent intervals to incorporate feedback from stakeholders as well as inclusion of latest developments. I am confident that revised curriculum has been framed after intense deliberations and would serve as a valuable resource to experts and student community.

I thank the Biotech Consortium India Limited for assisting DBT in this important exercise and compliment my colleague Dr. Suman Govil, Adviser, DBT for bringing out this publication.

(K. VijayRaghavan)
The biotechnology sector in India is extremely innovative and on the rise. Next few years are bound to see exponential growth in this sector. India is among the top 12 biotechnology destinations in the world and ranks third in the Asia-Pacific region. It is regarded as one of the most significant sectors in enhancing India’s global economic profile. India has been blessed with highly talented pool of students in biotechnology.

The National Biotechnology Development Strategy (2015 – 2020) and National Education Policy (2016) envisions a quality education system to produce graduates equipped with the knowledge, skills, attitudes and values that are required to lead a productive life and participate in the country’s development process. Improving employability in this sector is heavily dependent on the overall curriculum of the educational programs. Since last national course curriculum revision exercise was undertaken in 2008, it is necessary to update the current curriculum. In view of the scientific advancements taking place globally in the field of biotechnology, it was highly desirable to update the current course accordingly and modify it based on the needs of both research and industry. The Department of Biotechnology, Ministry of Science and Technology, Government of India through Biotech Consortium India Limited (BCIL), took the mandate to update the curriculum to keep abreast with the latest developments and to meet the needs of skilled manpower in rapidly advancing field of biotechnology. After continuous deliberations, discussions and several brainstorming sessions, the Core Committee and Subject-specific Committees comprising of experts in various fields of biotechnology arrived at revised and updated course curricula for 13 post-graduate programmes in biotechnology. Each course curriculum has about 90 – 100 credits comprising of core subjects, technology-based subjects, practicals and electives. These course curricula shall serve as model guidelines for academicians across the country for design and development of course curriculum in their institutions.

It is envisaged that the course curriculum revision exercise shall promote outcome-based education to meet not just the local and national manpower requirements, but also provide personal satisfaction and career potential for professionals with supporting pathways for their development.

In concluding this important task for human resource development, I thank my Co-chair Prof. K.K.Rao, fellow members of the Core Committee and Subject-Specific Committees for contributing towards this exercise. I would like to congratulate DBT for taking up this exercise and also BCIL for successfully coordinating it. I am confident that the present guidelines shall be extremely useful to the educators for imparting quality education for biotechnology in the country.

(Prof. Rakesh Bhatnagar)
Biotechnology in India has been a rapidly growing industry and has captured the essence of life beautifully. It is estimated that this industry will soon become a $100 billion entity. Universities are the platform of intellectual growth and course curriculum helps the students reach their goals which can be either research or industry. Thus, it is mandatory to keep updating the course curriculum regularly so that the students are able to take the most informed route to reach their goals.

The Department of Biotechnology, Ministry of Science and Technology, Government of India has been actively making numerous efforts to raise the biotechnology sector in India. Looking at this rapidly growing sector and the ever increasing gap of knowledge between university graduates and industry recruits led to the need for an updated course curricula. The responsibility for this task was entrusted to the Biotech Consortium India Limited, New Delhi to arrive at the best possible methods to update the current curricula keeping in mind the needs of students and industries. This exercise was previously undertaken in 2008 and hence this exercise was long overdue. The current updation exercise not only brings the course curricula at par with the current development in biotechnology, but also seeks to create a manpower and human resource capable of high order thinking and skills. Several inclusions like bioentrepreneurship and IPR will help the graduates to explore the $100 billion market, while other inclusions like Critical Analysis of Classical Papers and Project proposal and Presentation would help them augment the market to $200 billion.

This booklet is a result of several deliberations between the esteemed Core committee and the numerous subject specific committees. After several rounds of personal interactions and numerous discussions, we have tried to come up with the best possible course curricula for the universities to implement. In conclusion, I would like to thank and congratulate the Department of Biotechnology, Ministry of Science and Technology, Government of India for extending its support; BCIL for conducting this exercise efficiently; the core committee and all the experts in different committees for lending their valuable time and expertise to this noble cause. I hope this guideline curricula will enable the educators to impart best possible knowledge and learning experience to students.

(K. Krishnamurthy Rao)
Preface

Background

Promotion of Indian Biotechnology sector is high on policy agenda of Government of India. Biotechnology has also been recognized as one of the key priority sectors under ‘Make in India,’ 'Skill India' and 'Startup India' initiatives of Government of India, as it is one of sectors expected to contribute towards enterprise creation, innovation and economic growth. Department of Biotechnology (DBT), Ministry of Science and Technology, Government of India has immensely contributed to this dynamism through various policies and initiatives, establishment of innovation clusters, academia-industry partnerships, increasing capabilities for technology development, etc. The National Biotechnology Development Strategy (2015 – 2020) released by DBT provides a strategic roadmap for India's emergence as a global biotechnology innovation and manufacturing hub. It has also highlighted importance of human resource development and need for nurturing tailor-made human capital for advanced scientific research and entrepreneurship.

DBT has taken a number of initiatives aimed at integrated human resource development to evolve an ecosystem where scientists, innovators and future entrepreneurs can be nurtured. Keeping in mind requirement for trained manpower in various areas of Biotechnology, DBT initiated Post-Graduate Teaching Programme way back in 1985 with 5 universities which has expanded to 74 universities imparting M.Sc./M.Tech./M.V.Sc. degrees in general, agricultural, animal, food, environmental, industrial marine, medical, neuroscience and pharmaceutical biotechnology. 10 programmes are being phased out. These universities and institutes are provided liberal financial support towards strengthening of laboratory facilities, equipment, consumables, fellowships to students, dissertation grant per student etc. Post-Graduate Teaching Programme selects best students and trains them to join research or industry workforce contributing significantly to biotechnology workforce.

About the Course Curriculum Revision Exercise

Taking into cognizance the changing needs of the economy and to keep abreast with latest developments in the field of biotechnology, DBT proactively initiated revision of course curricula of Post-Graduate Programmes in biotechnology. The present exercise has been undertaken by Biotech Consortium India Limited (BCIL), New Delhi. Earlier exercise was carried out in 2008. The Course Curriculum Revision Exercise has been carried out for 13 Post-Graduate programmes in Biotechnology supported by DBT.

The revision of course curriculum of M.Sc. Biotechnology aims to address mismatch between 'knowledge' gained by students and appropriate skill set required for technology development and implementation including present contemporary needs of economy.

Methodology

A meticulous and structured approach has been adopted to accomplish the Course Curriculum Revision exercise.

BCIL had initiated the exercise with a review of literature of relevant national and international documents on curriculum design and planning for biotechnology programmes of premier national as well as international universities, guidelines by University Grants Commission, recent curricular guidelines released by Indian Council of Agricultural Research, Ministry of Health and Family Welfare and Indian Institute of Science Education & Research and other relevant research papers on curriculum development in peer-reviewed journals.
The findings of the literature review were adopted to design questionnaires for eliciting feedback from stakeholders of Biotechnology community i.e. academicians, scientists, industry representatives and students. Feedback was received from 165 experts and 20 students belonging to academic institutions, research organizations and industry regarding addition of advanced topics, deletion of elementary, redundant and overlapping topics, updation of laboratory practicals, re-adjustment of credit load, incorporating 'technology' component in the curriculum, among others. It was also suggested that re-orientation of curricula should be done keeping in view the needs of the industry.

**Strategic Approach**

A Core Committee along with 9 subject specific subcommittees comprising of 63 academicians, scientists and industry representatives were constituted to revise and update the curricula. The constitution of Core Committee is given at Annexure-1.

The salient recommendations identified from stakeholder survey were presented to the Committee. Several brainstorming discussion sessions were held for achieving the desired balance between the foundation courses, recent developments in biotechnology and updation needs identified during the stakeholder survey. Core Committee finalized broad contours for revising all the course curricula including M.Sc. Biotechnology. The guidelines set by the Core Committee were taken up by the subject specific committees for updating the respective curricula. The revised curriculum was vetted and finalized by the Core Committee.

**Course Curriculum Revision**

The members of Committee agreed that revised course curriculum should provide skill and outcome based education and help the students to gain domain knowledge, ability to design and interpret research experiments and acquire effective communication skills. The course curriculum has been re-designed accordingly to promote skill-based and outcome-based education. The revised course curriculum totals to 94 credits comprising of theory, practical, technology-based topics, electives and dissertation. Each course includes learning objectives, student learning outcomes, course plan (number of lectures/unit) and reference textbooks/resources. Theory and practical courses include relevant examples, case scenarios and tutorials for inculcating critical thinking against rote learning. Bridging courses in Chemistry, Physics, Mathematics and Statistics have been introduced. Content for Foundation courses Biochemistry, Cell and Molecular Biology, Genetics, Microbiology, Immunology and Genetic Engineering has been updated. Courses such as Critical Analysis of Classical Papers and Emerging Technologies have to develop problem-solving approach. New courses such as Bioentrepreneurship, Intellectual Property Rights, Biosafety and Bioethics, Bioinformatics, Research Methodology, Scientific Communication Skills, etc. with a view to provide holistic education. With importance of students being able to execute research projects independently, separate credits have been allotted for proposal preparation and presentation before initiating dissertation and also credits for dissertation have been increased accordingly.

We hope that model course curriculum shall serve as guidelines for academicians and researchers from different parts of the country for adoption in their institutions with modifications as per availability of expertise, infrastructure and specific needs.

We wish to put on record our sincere appreciation for constant guidance and encouragement received from Dr. K. VijayRaghavan, Secretary, DBT for bringing out this publication. We wish to acknowledge whole-hearted support of Core Committee and subject specific subcommittees members. Sincere thanks are due to Dr. Manoj Singh Rohilla, Scientist- D, DBT, Ms. Shweta for creative design, Mrs. Rita Bhatla, DBT and Shri. Dilip Joy, BCIL.
M.Sc. Biotechnology

<table>
<thead>
<tr>
<th>S.No.</th>
<th>Title</th>
<th>Credits</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td><strong>SEMESTER ONE</strong></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Biochemistry</td>
<td>3</td>
</tr>
<tr>
<td>2</td>
<td>Cell and Molecular Biology</td>
<td>3</td>
</tr>
<tr>
<td>3</td>
<td>Plant and Animal Biotechnology</td>
<td>3</td>
</tr>
<tr>
<td>4</td>
<td>Microbiology</td>
<td>2</td>
</tr>
<tr>
<td>5</td>
<td>Genetics</td>
<td>2</td>
</tr>
<tr>
<td>6</td>
<td>Basics of Mathematics and Statistics</td>
<td>2</td>
</tr>
<tr>
<td>7</td>
<td>Basics of Chemistry and Physics</td>
<td>2</td>
</tr>
<tr>
<td>8</td>
<td>Laboratory I: Biochemistry and Analytical Techniques</td>
<td>4</td>
</tr>
<tr>
<td>9</td>
<td>Laboratory II: Microbiology</td>
<td>2</td>
</tr>
<tr>
<td>10</td>
<td>Laboratory III: Plant and Animal Biotechnology</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td><strong>TOTAL</strong></td>
<td><strong>24</strong></td>
</tr>
<tr>
<td></td>
<td><strong>SEMESTER TWO</strong></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Genetic Engineering</td>
<td>3</td>
</tr>
<tr>
<td>2</td>
<td>Immunology</td>
<td>3</td>
</tr>
<tr>
<td>3</td>
<td>Bioinformatics</td>
<td>3</td>
</tr>
<tr>
<td>4</td>
<td>Genomics and Proteomics</td>
<td>2</td>
</tr>
<tr>
<td>5</td>
<td>Molecular Diagnostics</td>
<td>2</td>
</tr>
<tr>
<td>6</td>
<td>Research Methodology and Scientific Communication Skills</td>
<td>2</td>
</tr>
<tr>
<td>7</td>
<td>Elective I</td>
<td>2</td>
</tr>
<tr>
<td>8</td>
<td>Seminar</td>
<td>1</td>
</tr>
<tr>
<td>9</td>
<td>Laboratory IV: Molecular Biology and Genetic Engineering</td>
<td>4</td>
</tr>
<tr>
<td>10</td>
<td>Laboratory V: Immunology</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td><strong>TOTAL</strong></td>
<td><strong>24</strong></td>
</tr>
<tr>
<td></td>
<td><strong>SEMESTER THREE</strong></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Bioprocess Engineering and Technology</td>
<td>3</td>
</tr>
<tr>
<td>2</td>
<td>Emerging Technologies</td>
<td>2</td>
</tr>
<tr>
<td>3</td>
<td>Critical Analysis of Classical Papers</td>
<td>2</td>
</tr>
<tr>
<td>4</td>
<td>Bioentrepreneurship</td>
<td>2</td>
</tr>
<tr>
<td>5</td>
<td>Intellectual Property Rights, Biosafety and Bioethics</td>
<td>2</td>
</tr>
<tr>
<td>6</td>
<td>Project Proposal Preparation and Presentation</td>
<td>2</td>
</tr>
<tr>
<td>7</td>
<td>Seminar</td>
<td>1</td>
</tr>
<tr>
<td>8</td>
<td>Laboratory VI: Bioprocess Engineering and Technology</td>
<td>4</td>
</tr>
<tr>
<td>9</td>
<td>Laboratory VII: Bioinformatics</td>
<td>2</td>
</tr>
<tr>
<td>10</td>
<td>Dissertation</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td><strong>TOTAL</strong></td>
<td><strong>24</strong></td>
</tr>
<tr>
<td></td>
<td><strong>SEMESTER FOUR</strong></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Dissertation</td>
<td>20</td>
</tr>
<tr>
<td>2</td>
<td>Elective II</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td><strong>TOTAL</strong></td>
<td><strong>22</strong></td>
</tr>
<tr>
<td></td>
<td><strong>TOTAL CREDITS</strong></td>
<td><strong>94</strong></td>
</tr>
</tbody>
</table>

Recommended Electives:
# Semester One

## Biochemistry

**Credits**

<table>
<thead>
<tr>
<th>Unit I</th>
<th>Chemical basis of life</th>
<th>7 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Course Objectives</strong></td>
<td>Chemical basis of life: Miller-Urey experiment, abiotic formation of amino acid oligomers, composition of living matter; Water – properties of water, essential role of water for life on earth pH, buffer, maintenance of blood pH and pH of gastric juice, pH optima of different enzymes (pepsin, trypsin and alkaline phosphatase), ionization and hydrophobicity, emergent properties of biomolecules in water, biomolecular hierarchy, macromolecules, molecular assemblies.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit II</th>
<th>Protein structure</th>
<th>4 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Course Objectives</strong></td>
<td>Structure-function relationships: amino acids – structure and functional group properties, peptides and covalent structure of proteins, elucidation of primary and higher order structures, Ramachandran plot, evolution of protein structure, protein degradation and introduction to molecular pathways controlling protein degradation, structure-function relationships in model proteins like ribonuclease A, myoglobin, hemoglobin, chymotrypsin etc.; basic principles of protein purification; tools to characterize expressed proteins; Protein folding: Anfinsen's Dogma, Levinthal paradox, cooperativity in protein folding, free energy landscape of protein folding and pathways of protein folding, molten globule state, chaperons, diseases associated with protein folding, introduction to molecular dynamic simulation.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit III</th>
<th>Enzyme kinetics</th>
<th>5 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Course Objectives</strong></td>
<td>Enzyme catalysis – general principles of catalysis; quantitation of enzyme activity and efficiency; enzyme characterization and Michaelis-Menten kinetics; relevance of enzymes in metabolic regulation, activation, inhibition and covalent modification; single substrate enzymes; concept of catalytic antibodies; catalytic strategies with specific examples of proteases, carbonic anhydrases, restriction enzymes and nucleoside monophosphate kinase; regulatory strategies with specific example of hemoglobin; isozymes; role of covalent modification in enzymatic activity; zymogens.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit IV</th>
<th>Glycobiology</th>
<th>2 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Course Objectives</strong></td>
<td>Sugars - mono, di, and polysaccharides with specific reference to glycogen, amylase and cellulose, glycosylation of other biomolecules - glycoproteins and glycolipids; lipids - structure and properties of important members of storage and membrane lipids; lipoproteins.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit V</th>
<th>Structure and functions of DNA &amp; RNA and lipids</th>
<th>3 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Course Objectives</strong></td>
<td>Self-assembly of lipids, micelle, biomembrane organization - sidedness and function; membrane bound proteins - structure, properties and function; transport phenomena; nucleosides, nucleotides, nucleic acids - structure, a historical perspective leading up to the proposition of DNA double helical structure; difference in RNA and DNA structure and their importance in evolution of DNA as the genetic material.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit VI</th>
<th>Bioenergetics</th>
<th>8 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Course Objectives</strong></td>
<td>Bioenergetics-basic principles; equilibria and concept of free energy; coupled interconnecting reactions in metabolism; oxidation of carbon fuels; recurring motifs in metabolism; Introduction to GPCR, Inositol/DAG//PKC and Ca++ signaling pathways;</td>
<td></td>
</tr>
</tbody>
</table>
glycolysis and gluconeogenesis; reciprocal regulations and non-carbohydrate sources of glucose; Citric acid cycle, entry to citric acid cycle, citric acid cycle as a source of biosynthetic precursors; Oxidative phosphorylation; importance of electron transfer in oxidative phosphorylation; F1-F0 ATP Synthase; shuttles across mitochondria; regulation of oxidative phosphorylation; Photosynthesis – chloroplasts and two photosystems; proton gradient across thylakoid membrane; Calvin cycle and pentose phosphate pathway; glycogen metabolism, reciprocal control of glycogen synthesis and breakdown, roles of epinephrine and glucagon and insulin in glycogen metabolism; Fatty acid metabolism; protein turnover and amino acid catabolism; nucleotide biosynthesis; biosynthesis of membrane lipids and sterols with specific emphasis on cholesterol metabolism and mevalonate pathway; elucidation of metabolic pathways; logic and integration of central metabolism; entry/ exit of various biomolecules from central pathways; principles of metabolic regulation; steps for regulation.

Calvin cycle and pentose phosphate pathway; glycogen metabolism, reciprocal control of glycogen synthesis and breakdown, roles of epinephrine and glucagon and insulin in glycogen metabolism; Fatty acid metabolism; protein turnover and amino acid catabolism; nucleotide biosynthesis; biosynthesis of membrane lipids and sterols with specific emphasis on cholesterol metabolism and mevalonate pathway; elucidation of metabolic pathways; logic and integration of central metabolism; entry/ exit of various biomolecules from central pathways; principles of metabolic regulation; steps for regulation; target of rapamycin (TOR) & Autophagy regulation in relation to C & N metabolism, starvation responses and insulin signaling.

Recommended Textbooks and References:
Chromatin organization - histone and DNA interactome: structure and assembly of eukaryotic and prokaryotic DNA polymerases, DNA-replication, repair and recombination; chromatin control: gene transcription and silencing by chromatin-Writers, Readers and –Erasers; Transcriptional control: Structure and assembly of eukaryotic and prokaryotic RNA Polymerases, promoters and enhancers, transcription factors as activators and repressors, transcriptional initiation, elongation and termination; post-transcriptional control: splicing and addition of cap and tail, mRNA flow through nuclear envelope into cytoplasm, breakdown of selective and specific mRNAs through interference by small non-coding RNAs (miRNAs and siRNAs), protein translation machinery, ribosomes-composition and assembly; universal genetic codes, degeneracy of codons, Wobble hypothesis; Iso-accepting tRNA; mechanism of initiation, elongation and termination; co- and post-translational modifications, mitochondrial genetic code translation product cleavage, modification and activation.

Molecular mechanisms of membrane transport, nuclear transport, transport across mitochondria and chloroplasts; intracellular vesicular trafficking from endoplasmic reticulum through Golgi apparatus to lysosomes/cell exterior.

Cell cycle and its regulation; cell division: mitosis, meiosis and cytokinesis; cell differentiation: stem cells, their differentiation into different cell types and organization into specialized tissues; cell-ECM and cell-cell interactions; cell receptors and trans-membrane signalling; cell motility and migration; cell death: different modes of cell death and their regulation.

Isolation of cells and basics of cell culture; observing cells under a microscope, different types of microscopy; analyzing and manipulating DNA, RNA and proteins.

Mutations, proto-oncogenes, oncogenes and tumour suppressor genes, physical, chemical and biological mutagens; types of mutations; intra-genic and inter-genic suppression; transpositions- transposable genetic elements in prokaryotes and eukaryotes, role of transposons in genome; viral and cellular oncogenes; tumor suppressor genes; structure, function and mechanism of action; activation and suppression of tumor suppressor genes; oncogenes as transcriptional activators.

Recommended Textbooks and References:
**Course Objectives**
The objectives of this course are to introduce students to the principles, practices and application of animal biotechnology, plant tissue culture, plant and animal genomics, genetic transformation and molecular breeding of plants and animals.

**Student Learning Outcomes**
Students should be able to gain fundamental knowledge in animal and plant biotechnology and their applications.

<table>
<thead>
<tr>
<th>Unit</th>
<th>Topic</th>
<th>Lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Unit I</strong></td>
<td><strong>Plant tissue culture and animal cell culture</strong></td>
<td>10 lectures</td>
</tr>
<tr>
<td></td>
<td>Plant tissue culture: historical perspective; totipotency; organogenesis; Somatic embryogenesis; establishment of cultures – callus culture, cell suspension culture, media preparation – nutrients and plant hormones; sterilization techniques; applications of tissue culture - micropropagation; somaclonal variation; androgenesis and its applications in genetics and plant breeding; germplasm conservation and cryopreservation; synthetic seed production; protoplast culture and somatic hybridization - protoplast isolation; culture and usage; somatic hybridization - methods and applications; cybrids and somatic cell genetics; plant cell cultures for secondary metabolite production. Animal cell culture: brief history of animal cell culture; cell culture media and reagents; culture of mammalian cells, tissues and organs; primary culture, secondary culture, continuous cell lines, suspension cultures; application of animal cell culture for virus isolation and <em>in vitro</em> testing of drugs, testing of toxicity of environmental pollutants in cell culture, application of cell culture technology in production of human and animal viral vaccines and pharmaceutical proteins.</td>
<td></td>
</tr>
<tr>
<td><strong>Unit II</strong></td>
<td><strong>Plant genetic manipulation</strong></td>
<td>10 lectures</td>
</tr>
<tr>
<td></td>
<td>Genetic engineering: <em>Agrobacterium</em>-plant interaction; virulence; Ti and Ri plasmids; opines and their significance; T-DNA transfer; disarmed Ti plasmid; Genetic transformation - <em>Agrobacterium</em>-mediated gene delivery; cointegrate and binary vectors and their utility; direct gene transfer - PEG-mediated, electroporation, particle bombardment and alternative methods; screenable and selectable markers; characterization of transgenics; chloroplast transformation; marker-free methodologies; advanced methodologies - cisgenesis, intragenesis and genome editing; molecular pharming - concept of plants as biofactories, production of industrial enzymes and pharmaceutically important compounds.</td>
<td></td>
</tr>
<tr>
<td><strong>Unit III</strong></td>
<td><strong>Animal reproductive biotechnology and vaccinology</strong></td>
<td>8 lectures</td>
</tr>
<tr>
<td></td>
<td>Animal reproductive biotechnology: structure of sperms and ovum; cryopreservation of sperms and ova of livestock; artificial insemination; super ovulation, embryo recovery and <em>in vitro</em> fertilization; culture of embryos; cryopreservation of embryos; embryo transfer technology; transgenic manipulation of animal embryos; applications of transgenic animal technology; animal cloning - basic concept, cloning for conservation for conservation endangered species; Vaccinology: history of development of vaccines, introduction to the concept of vaccines, conventional methods of animal vaccine production, recombinant approaches to vaccine production, modern vaccines.</td>
<td></td>
</tr>
<tr>
<td><strong>Unit IV</strong></td>
<td><strong>Plant and animal genomics</strong></td>
<td>4 lectures</td>
</tr>
<tr>
<td></td>
<td>Overview of genomics – definition, complexity and classification; need for genomics level analysis; methods of analyzing genome at various levels – DNA, RNA, protein, metabolites and phenotype; genome projects and bioinformatics resources for genome research – databases; overview of forward and reverse genetics for assigning function for genes.</td>
<td></td>
</tr>
</tbody>
</table>
Molecular markers - hybridization and PCR based markers RFLP, RAPD, STS, SSR, AFLP, SNP markers; DNA fingerprinting-principles and applications; introduction to mapping of genes/QTLs; marker-assisted selection - strategies for introducing genes of biotic and abiotic stress resistance in plants; genetic basis for disease resistance in animals; molecular diagnostics of pathogens in plants and animals; detection of meat adulteration using DNA based methods.

Recommended Textbooks and References:

Course Objectives
The objectives of this course are to introduce field of microbiology with special emphasis on microbial diversity, morphology, physiology and nutrition; methods for control of microbes and host-microbe interactions.

Student Learning Outcomes
Students should be able to:
• Identify major categories of microorganisms and analyze their classification, diversity, and ubiquity;
• Identify and demonstrate structural, physiological, genetic similarities and differences of major categories of microorganisms;
• Identify and demonstrate how to control microbial growth;
• Demonstrate and evaluate interactions between microbes, hosts and environment.

Unit I
Microbial characteristics
6 lectures
Introduction to microbiology and microbes, history & scope of microbiology, morphology, structure, growth and nutrition of bacteria, bacterial growth curve, bacterial culture methods; bacterial genetics: mutation and recombination in bacteria, plasmids, transformation, transduction and conjugation; antimicrobial resistance.

Unit II
Microbial diversity
9 lectures
Microbial taxonomy and evolution of diversity, classification of microorganisms, criteria for classification; classification of bacteria; Cyanobacteria, acetic acid bacteria, Pseudomonads, lactic and propionic acid bacteria, endospore forming bacteria,
**Mycobacteria and Mycoplasma. Archaea: Halophiles, Methanogens, Hyperthermophilic archae, Thermoplasm; eukarya: algae, fungi, slime molds and protozoa; extremophiles and unculturable microbes.**

**Unit III**
**Control of microorganisms**
3 lectures

Sterilization, disinfection and antisepsis: physical and chemical methods for control of microorganisms, antibiotics, antiviral and antifungal drugs, biological control of microorganisms.

**Unit IV**
**Virology**
5 lectures

Virus and bacteriophages, general properties of viruses, viral structure, taxonomy of virus, viral replication, cultivation and identification of viruses; sub-viral particles – viroids and prions.

**Unit V**
**Host-microbes interaction**
5 lectures

Host-pathogen interaction, ecological impact of microbes; symbiosis (Nitrogen fixation and ruminant symbiosis); microbes and nutrient cycles; microbial communication system; bacterial quorum sensing; microbial fuel cells; prebiotics and probiotics.

**Recommended Textbooks and References:**

---

**Genetics**

**Credits**
2

The objectives of this course are to take students through basics of genetics and classical genetics covering prokaryotic/phage genetics to yeast and higher eukaryotic domains. On covering all classical concepts of Mendelian genetics across these life-forms, students will be exposed to concepts of population genetics, quantitative genetics encompassing complex traits, clinical genetics and genetics of evolution.

**Course Objectives**

**Student Learning Outcomes**

On successful completion of this course, student will be able:

- Describe fundamental molecular principles of genetics;
- Understand relationship between phenotype and genotype in human genetic traits;
- Describe the basics of genetic mapping;
- Understand how gene expression is regulated.

**Unit I**
**Genetics of bacteria and bacteriophages**
10 lectures

Concept of a gene in pre-DNA era; mapping of genes in bacterial and phage chromosomes by classical genetic crosses; fine structure analysis of a gene; genetic complementation and other genetic crosses using phenotypic markers; phenotype to genotype connectivity prior to DNA-based understanding of gene.

**Unit II**
**Yeast genetics**
6 lectures

Meiotic crosses, tetrad analyses, non-Mendelian and Mendelian ratios, gene conversion, models of genetic recombination, yeast mating type switch; dominant and recessive genes/mutations, suppressor or modifier screens, complementation groups, transposon mutagenesis, synthetic lethality, genetic epistasis.
Basics of Mathematics and Statistics

Unit I
Algebra
6 lectures
Linear equations, functions: slopes-intercepts, forms of two-variable linear equations; constructing linear models in biological systems; quadratic equations (solving, graphing, features of, interpreting quadratic models etc.), introduction to polynomials, graphs of binomials and polynomials; Symmetry of polynomial functions, basics of trigonometric functions, Pythagorean theory, graphing and constructing sinusoidal functions, imaginary numbers, complex numbers, adding-subtracting-multiplying complex numbers, basics of vectors, introduction to matrices.

Unit II
Calculus
4 lectures
Differential calculus (limits, derivatives), integral calculus (integrals, sequences and series etc.).

Unit III
Drosophila genetics as a model of higher eukaryotes
4 lectures
Monohybrid & dihybrid crosses, back-crosses, test-crosses, analyses of autosomal and sex linkages, screening of mutations based on phenotypes and mapping the same, hypomorphy, genetic mosaics, genetic epistasis in context of developmental mechanism.

Unit IV
Population genetics and genetics of evolution
4 lectures
Introduction to the elements of population genetics: genetic variation, genetic drift, neutral evolution; mutation selection, balancing selection, Fishers theorem, Hardy-Weinberg equilibrium, linkage disequilibrium; in-breeding depression & mating systems; population bottlenecks, migrations, Bayesian statistics; adaptive landscape, spatial variation & genetic fitness.

Unit V
Quantitative genetics of complex traits (QTLs)
2 lectures
Complex traits, mapping QTLs, yeast genomics to understand biology of QTLs.

Unit VI
Plant genetics
2 lectures
Laws of segregation in plant crosses, inbreeding, selfing, heterosis, maintenance of genetic purity, gene pyramiding.

Recommended Textbooks and References:

Course Objectives
The objective of this course is to give conceptual exposure of essential contents of mathematics and statistics to students.

Student Learning Outcomes
On completion of this course, students should be able to:
• Gain broad understanding in mathematics and statistics;
• Recognize importance and value of mathematical and statistical thinking, training, and approach to problem solving, on a diverse variety of disciplines.
Population dynamics; oscillations, circadian rhythms, developmental patterns, symmetry in biological systems, fractal geometries, size-limits & scaling in biology, modeling chemical reaction networks and metabolic networks.

Probability: counting, conditional probability, discrete and continuous random variables; Error propagation; Populations and samples, expectation, parametric tests of statistical significance, nonparametric hypothesis tests, linear regression, correlation & causality, analysis of variance, factorial experiment design.

Recommended Textbooks and References:

Course Objectives
The objectives of this course are to cover all essentials required to appreciate physico-chemical principles underlying biological processes.

Student Learning Outcomes
Students should be able to have a firm foundation in fundamentals and application of current chemical and physical scientific theories.

Physical quantities and their dynamics: definitions and dimensions; vectors & scalars, displacement, velocity, acceleration, kinematic formulas, angular momentum, torque etc. force, power, work, energy (kinetic & potential/electric charge separation, electromagnetic spectrum, photons etc.); springs & Hooke's laws; elastic and inelastic collisions; Newton's law of motions (centripetal and centrifugal forces etc.); simple harmonic motions, mechanical waves, Doppler effect, wave interference, amplitude, period, frequency & wavelength; diffusion, dissipation, random walks, and directed motions in biological systems; low Reynolds number - world of Biology, buoyant forces, Bernoulli's equation, viscosity, turbulence, surface tension, adhesion; laws of thermodynamics: Maxwell Boltzmann distribution, conduction, convection and radiation, internal energy, entropy, temperature and free energy, Maxwell's demon (entropic forces at work in biology, chemical assemblies, self-assembled systems, role of ATP); Coulomb's law, conductors and insulators, electric potential energy of charges, nerve impulses, voltage gated channels, ionic conductance; Ohms law (basic electrical quantities: current, voltage & power), electrolyte conductivity, capacitors and capacitance, dielectrics; various machines in biology i.e. enzymes, allostery and molecular motors (molecules to cells and organisms).

Basic constituents of matter - elements, atoms, isotopes, atomic weights, atomic numbers, basics of mass spectrometry, molecules, Avogadro number, molarity, gas constant, molecular weights, structural and molecular formulae, ions and polyatomic
12 lectures:
10 hours teaching + 2 hours tutorials

ions; chemical reactions, reaction stoichiometry, rates of reaction, rate constants, order of reactions, Arrhenious equation, Maxwell Boltzmann distributions, rate-determining steps, catalysis, free-energy, entropy and enthalpy changes during reactions; kinetic versus thermodynamic controls of a reaction, reaction equilibrium (equilibrium constant); light and matter interactions (optical spectroscopy, fluorescence, bioluminescence, paramagnetism and diamagnetism, photoelectron spectroscopy; chemical bonds (ionic, covalent, Van der Walls forces); electronegativity, polarity; VSEPR theory and molecular geometry, dipole moment, orbital hybridizations; states of matter - vapor pressure, phase diagrams, surface tension, boiling and melting points, solubility, capillary action, suspensions, colloids and solutions; acids, bases and pH - Arrhenious theory, pH, ionic product of water, weak acids and bases, conjugate acid-base pairs, buffers and buffering action etc; chemical thermodynamics - internal energy, heat and temperature, enthalpy (bond enthalpy and reaction enthalpy), entropy, Gibbs free energy of ATP driven reactions, spontaneity versus driven reactions in biology; redox reactions and electrochemistry - oxidation-reduction reactions, standard cell potentials, Nernst equation, resting membrane potentials, electron transport chains (ETC) in biology, coupling of oxidative phosphorylations to ETC; theories of ATP production and dissipation across biological membranes; bond rotations and molecular conformations - Newman projections, conformational analysis of alkanes, alkenes and alkynes; functional groups, optically asymmetric carbon centers, amino acids, proteins, rotational freedoms in polypeptide backbone (Ramachandran plot).

Recommended Textbooks and References:

Course Objectives
The objective of this laboratory course is to introduce students to experiments in biochemistry. The course is designed to teach students the utility of set of experimental methods in biochemistry in a problem oriented manner.

Student Learning Outcomes
On completion of this course, students should be able to:
• To elaborate concepts of biochemistry with easy to run experiments;
• To familiarize with basic laboratory instruments and understand the principle of measurements using those instruments with experiments in biochemistry.

Syllabus
1. Preparing various stock solutions and working solutions that will be needed for the course.

2. To prepare an Acetic-Na Acetate Buffer and validate the Henderson-Hasselbach equation.
3. To determine an unknown protein concentration by plotting a standard graph of BSA using UV-Vis Spectrophotometer and validating the Beer- Lambert's Law.
4. Titration of Amino Acids and separation of aliphatic, aromatic and polar amino acids by thin layer chromatography.
5. Purification and characterization of an enzyme from a recombinant source (such as Alkaline Phosphatase or Lactate Dehydrogenase or any enzyme of the institution's choice).
   a) Preparation of cell-free lysates
   b) Ammonium Sulfate precipitation
   c) Ion-exchange Chromatography
   d) Gel Filtration
   e) Affinity Chromatography
   f) Dialysis of the purified protein solution against 60% glycerol as a demonstration of storage method
   g) Generating a Purification Table (protein concentration, amount of total protein; Computing specific activity of the enzyme preparation at each stage of purification)
   h) Assessing purity of samples from each step of purification by SDS-PAGE Gel Electrophoresis
   i) Enzyme Kinetic Parameters: Km, Vmax and Kcat.
6. Experimental verification that absorption at OD$_{260}$ is more for denatured DNA as compared to native double stranded DNA. reversal of the same following DNA renaturation. Kinetics of DNA renaturation as a function of DNA size.
7. Identification of an unknown sample as DNA, RNA or protein using available laboratory tools. (Optional Experiments)

---

**Course Objectives**

The objective of this laboratory course is to provide practical skills on basic microbiological techniques.

**Student Learning Outcomes**

Students should be able to:

- Isolate, characterize and identify common bacterial organisms;
- Determine bacterial load of different samples;
- Perform antimicrobial sensitivity tests;
- Preserve bacterial cultures.

---

**Syllabus**

1. Sterilization, disinfection and safety in microbiological laboratory.
2. Preparation of media for cultivation of bacteria.
3. Isolation of bacteria in pure culture by streak plate method.
4. Study of colony and growth characteristics of some common bacteria: *Bacillus, E. coli, Staphylococcus, Streptococcus, etc.*
5. Preparation of bacterial smear and Gram's staining.
7. Antimicrobial sensitivity test and demonstration of drug resistance.
8. Maintenance of stock cultures: slants, stabs and glycerol stock cultures
9. Determination of phenol co-efficient of antimicrobial agents.
10. Determination of Minimum Inhibitory Concentration (MIC)
11. Isolation and identification of bacteria from soil/water samples.

**Recommended Textbooks and References:**

3. Tille, P. M., & Forbes, B. A. *Bailey & Scott's Diagnostic Microbiology*.

---

**Course Objectives**

The objectives of this course are to provide hands-on training in basic experiments of plant and animal biotechnology.

**Student Learning Outcomes**

On completion of course, students should be able to gain basic skills in plant and animal biotechnology.

---

**Syllabus**

**Plant Biotechnology**

1. Prepare culture media with various supplements for plant tissue culture.
2. Prepare explants of *Valleriana wallichii* for inoculation under aseptic conditions.
3. Attempt in vitro andro and gynogenesis in plants (*Datura stramonium*).
4. Isolate plant protoplast by enzymatic and mechanical methods and attempt fusion by PEG (available material).
5. Culture *Agrobacterium tumefaciens* and attempt transformation of any dicot species.
6. Generate an RAPD and ISSR profile of *Eremurus persicus* and *Valleriana wallichii*.
7. Prepare karyotypes and study the morphology of somatic chromosomes of *Allium cepa*, *A. sativum*, *A. tuberosum* and compare them on the basis of karyotypes.
8. Pollen mother cell meiosis and recombination index of select species (one achiasmate, and the other chiasmate) and correlate with generation of variation.
9. Undertake plant genomic DNA isolation by CTAB method and its quantitation by visual as well as spectrophotometric methods.
10. Perform PCR amplification of ‘n’ number of genotypes of a species for studying the genetic variation among the individuals of a species using random primers.
11. Study genetic fingerprinting profiles of plants and calculate polymorphic information content.

**Animal Biotechnology**

1. Count cells of an animal tissue and check their viability.
2. Prepare culture media with various supplements for plant and animal tissue culture.
3. Prepare single cell suspension from spleen and thymus.
5. Chromosome preparations from cultured animal cells.
6. Isolate DNA from animal tissue by SDS method.
7. Attempt animal cell fusion using PEG.
### Course Objectives

The objectives of this course are to teach students with various approaches to conducting genetic engineering and their applications in biological research as well as in biotechnology industries. Genetic engineering is a technology that has been developed based on our fundamental understanding of the principles of molecular biology and this is reflected in the contents of this course.

### Student Learning Outcomes

Given the impact of genetic engineering in modern society, the students should be endowed with strong theoretical knowledge of this technology. In conjunction with the practicals in molecular biology & genetic engineering, the students should be able to take up biological research as well as placement in the relevant biotech industry.

### Unit I
**Introduction and tools for genetic engineering**

- Impact of genetic engineering in modern society; general requirements for performing a genetic engineering experiment; restriction endonucleases and methylases; DNA ligase, Klenow enzyme, T4 DNA polymerase, polynucleotide kinase, alkaline phosphatase; cohesive and blunt end ligation; linkers; adaptors; homopolymeric tailing; labelling of DNA: nick translation, random priming, radioactive and non-radioactive probes, hybridization techniques: northern, southern, south-western and far-western and colony hybridization, fluorescence *in situ* hybridization.

### Unit II
**Different types of vectors**

- Plasmids; Bacteriophages; M13 mp vectors; PUC19 and Bluescript vectors, hagemids; Lambda vectors; Insertion and Replacement vectors; Cosmids; Artificial chromosome vectors (YACs; BACs); Principles for maximizing gene expression expression vectors; pMal; GST; pET-based vectors; Protein purification; His-tag; GST-tag; MBP-tag etc.; Intein-based vectors; Inclusion bodies; methodologies to reduce formation of inclusion bodies; mammalian expression and replicating vectors; Baculovirus and *Pichia* vectors system, plant based vectors, Ti and Ri as vectors, yeast vectors, shuttle vectors.

### Unit III
**Different types of PCR techniques**

- Principles of PCR: primer design; fidelity of thermostable enzymes; DNA polymerases; types of PCR – multiplex, nested; reverse-transcription PCR, real time PCR, touchdown PCR, hot start PCR, colony PCR, asymmetric PCR, cloning of PCR products; T-vectors; proof reading enzymes; PCR based site specific mutagenesis; PCR in molecular diagnostics; viral and bacterial detection; sequencing methods; enzymatic DNA sequencing; chemical sequencing of DNA; automated DNA sequencing; RNA sequencing; chemical synthesis of oligonucleotides; mutation detection: SSCP, DGGE, RFLP.

### Unit IV
**Gene manipulation and protein-DNA interaction**

- Insertion of foreign DNA into host cells; transformation, electroporation, transfection; construction of libraries; isolation of mRNA and total RNA; reverse transcriptase and cDNA synthesis; cDNA and genomic libraries; construction of microarrays – genomic arrays, cDNA arrays and oligo arrays; study of protein-DNA interactions: electrophoretic mobility shift assay; DNase footprinting; methyl interference assay, chromatin immunoprecipitation; protein-protein interactions using yeast two-hybrid system; phage display.

### Unit V
**Gene silencing and genome editing technologies**

- Gene silencing techniques; introduction to siRNA; siRNA technology; Micro RNA; construction of siRNA vectors; principle and application of gene silencing; gene knockouts and gene therapy; creation of transgenic plants; debate over GM crops; introduction to methods of genetic manipulation in different model systems e.g. fruit flies.
(Drosophila), worms (C. elegans), frogs (Xenopus), fish (zebra fish) and chick; Transgenics - gene replacement; gene targeting; creation of transgenic and knock-out mice; disease model; introduction to genome editing by CRISPR-CAS with specific emphasis on Chinese and American clinical trials.

Recommended Textbooks and References:

4. Selected papers from scientific journals, particularly Nature & Science.
5. Technical Literature from Stratagene, Promega, Novagen, New England Biolab etc.

Course Objectives

The objectives of this course are to learn about structural features of components of immune system as well as their function. The major emphasis of this course will be on development of immune system and mechanisms by which our body elicits immune response. This will be imperative for students as it will help them to predict about nature of immune response that develops against bacterial, viral or parasitic infection, and prove it by designing new experiments.

Student Learning Outcomes

On completion of this course, students should be able to:
- Evaluate usefulness of immunology in different pharmaceutical companies;
- Identify proper research lab working in area of their own interests;
- Apply their knowledge and design immunological experiments to demonstrate innate, humoral or cytotoxic T lymphocyte responses and figure out kind of immune responses in the setting of infection (viral or bacterial).

Unit I

Immunology: fundamental concepts and overview of the immune system
5 lectures

Components of innate and acquired immunity; phagocytosis; complement and inflammatory responses; pathogen recognition receptors (PRR) and pathogen associated molecular pattern (PAMP); innate immune response; mucosal immunity; antigens: immunogens, haptens; Major Histocompatibility Complex: MHC genes, MHC and immune responsiveness and disease susceptibility, Organs of immune system, primary and secondary lymphoid organs.

Unit II

Immune responses generated by B and T lymphocytes
8 lectures

Immunoglobulins - basic structure, classes & subclasses of immunoglobulins, antigenic determinants; multigene organization of immunoglobulin genes; B-cell receptor; Immunoglobulin superfamily; principles of cell signaling; basis of self & non-self discrimination; kinetics of immune response, memory; B cell maturation, activation and differentiation; generation of antibody diversity; T-cell maturation, activation and differentiation and T-cell receptors; functional T Cell subsets; cell-mediated immune responses, ADCC; cytokines: properties, receptors and therapeutic uses; antigen processing and presentation- endogenous antigens, exogenous antigens, non-peptide bacterial antigens and super-antigens; cell-cell co-operation, Hapten-carrier system.

Unit III

Antigen-antibody interactions
6 lectures

Precipitation, agglutination and complement mediated immune reactions; advanced immunological techniques: RIA, ELISA, Western blotting, ELISPOT assay, immunofluorescence microscopy, flow cytometry and immunoelectron microscopy; surface plasmon resonance, biosensor assays for assessing ligand –receptor interaction; CMI techniques: lymphoproliferation assay, mixed lymphocyte reaction, cell cytotoxicity assays, apoptosis, microarrays, transgenic mice, gene knock outs.
Active and passive immunization; live, killed, attenuated, subunit vaccines; vaccine technology: role and properties of adjuvants, recombinant DNA and protein based vaccines, plant-based vaccines, reverse vaccinology; peptide vaccines, conjugate vaccines; antibody genes and antibody engineering: chimeric, generation of monoclonal antibodies, hybrid monoclonal antibodies; catalytic antibodies and generation of immunoglobulin gene libraries, idiotypic vaccines and marker vaccines, viral-like particles (VLPs), dendritic cell based vaccines, vaccine against cancer, T cell based vaccine, edible vaccine and therapeutic vaccine.

Immunity to infection: bacteria, viral, fungal and parasitic infections (with examples from each group); hypersensitivity: Type I-IV; autoimmunity; types of autoimmune diseases; mechanism and role of CD4+ T cells; MHC and TCR in autoimmunity; treatment of autoimmune diseases; transplantation: immunological basis of graft rejection; clinical transplantation and immunosuppressive therapy; tumor immunology: tumor antigens; immune response to tumors and tumor evasion of the immune system, cancer immunotherapy; immunodeficiency: primary immunodeficiencies, acquired or secondary immunodeficiencies, autoimmune disorder, anaphylactic shock, immunosenescence, immune exhaustion in chronic viral infection, immune tolerance, NK cells in chronic viral infection and malignancy.

Major histocompatibility complex genes and their role in autoimmune and infectious diseases, HLA typing, human major histocompatibility complex (MHC), Complement genes of the human major histocompatibility complex: implication for linkage disequilibrium and disease associations, genetic studies of rheumatoid arthritis, systemic lupus erythematosus and multiple sclerosis, genetics of human immunoglobulin, immunogenetics of spontaneous control of HIV, KIR complex.

Recommended Textbooks and References:

Course Objectives
The objectives of this course are to provide theory and practical experience of the use of common computational tools and databases which facilitate investigation of molecular biology and evolution-related concepts.

Student Learning Outcomes
Student should be able to:
- Develop an understanding of basic theory of these computational tools;
- Gain working knowledge of these computational tools and methods;
- Appreciate their relevance for investigating specific contemporary biological questions;
- Critically analyse and interpret results of their study.
# Unit I
**Bioinformatics basics**

5 lectures

- Bioinformatics basics: Computers in biology and medicine; Introduction to Unix and Linux systems and basic commands; Database concepts; Protein and nucleic acid databases; Structural databases; Biological XML DTD’s; pattern matching algorithm basics; databases and search tools: biological background for sequence analysis; Identification of protein sequence from DNA sequence; searching of databases similar sequence; NCBI; publicly available tools; resources at EBI; resources on web; database mining tools.

# Unit II
**DNA sequence analysis**

5 lectures

- DNA sequence analysis: gene bank sequence database; submitting DNA sequences to databases and database searching; sequence alignment; pairwise alignment techniques; motif discovery and gene prediction; local structural variants of DNA, their relevance in molecular level processes, and their identification; assembly of data from genome sequencing.

# Unit III
**Multiple sequence analysis**

5 lectures

- Multiple sequence analysis; multiple sequence alignment; flexible sequence similarity searching with the FASTA3 program package; use of CLUSTALW and CLUSTALX for multiple sequence alignment; submitting DNA protein sequence to databases: where and how to submit, SEQUIN, genome centres; submitting aligned sets of sequences, updating submitted sequences, methods of phylogenetic analysis.

# Unit IV
**Protein modelling**

5 lectures

- Protein modelling: introduction; force field methods; energy, buried and exposed residues; side chains and neighbours; fixed regions; hydrogen bonds; mapping properties onto surfaces; fitting monomers; RMS fit of conformers; assigning secondary structures; sequence alignment- methods, evaluation, scoring; protein completion: backbone construction and side chain addition; small peptide methodology; software accessibility; building peptides; protein displays; substructure manipulations, annealing.

# Unit V
**Protein structure prediction and virtual library**

6 lectures

- Protein structure prediction: protein folding and model generation; secondary structure prediction; analyzing secondary structures; protein loop searching; loop generating methods; homology modelling: potential applications, description, methodology, homologous sequence identification; align structures, align model sequence; construction of variable and conserved regions; threading techniques; topology fingerprint approach for prediction; evaluation of alternate models; structure prediction on a mystery sequence; structure aided sequence techniques of structure prediction; structural profiles, alignment algorithms, mutation tables, prediction, validation, sequence based methods of structure prediction, prediction using inverse folding, fold prediction; significance analysis, scoring techniques, sequence-sequence scoring; protein function prediction; elements of in silico drug design; Virtual library: Searching PubMed, current content, science citation index and current awareness services, electronic journals, grants and funding information.

---

**Recommended Textbooks and References:**

Course Objectives
The objectives of this course is to provide introductory knowledge concerning genomics, proteomics and their applications.

Student Learning Outcomes
Students should be able to acquire knowledge and understanding of fundamentals of genomics and proteomics, transcriptomics and metabolomics and their applications in various applied areas of biology.

Unit I
Basics of genomics and proteomics
3 lectures

Brief overview of prokaryotic and eukaryotic genome organization; extra-chromosomal DNA: bacterial plasmids, mitochondria and chloroplast.

Unit II
Genome mapping
4 lectures

Genetic and physical maps; markers for genetic mapping; methods and techniques used for gene mapping, physical mapping, linkage analysis, cytogenetic techniques, FISH technique in gene mapping, somatic cell hybridization, radiation hybrid maps, in situ hybridization, comparative gene mapping.

Unit III
Genome sequencing projects
3 lectures

Human Genome Project, genome sequencing projects for microbes, plants and animals, accessing and retrieving genome project information from the web.

Unit IV
Comparative genomics
5 lectures

Identification and classification of organisms using molecular markers- 16S rRNA typing/sequencing, SNPs; use of genomes to understand evolution of eukaryotes, track emerging diseases and design new drugs; determining gene location in genome sequence.

Unit V
Proteomics
5 lectures

Aims, strategies and challenges in proteomics; proteomics technologies: 2D-PAGE, isoelectric focusing, mass spectrometry, MALDI-TOF, yeast 2-hybrid system, proteome databases.

Unit VI
Functional genomics and proteomics
8 lectures

Transcriptome analysis for identification and functional annotation of gene, Contig assembly, chromosome walking and characterization of chromosomes, mining functional genes in genome, gene function- forward and reverse genetics, gene ethics; protein-protein and protein-DNA interactions; protein chips and functional proteomics; clinical and biomedical applications of proteomics; introduction to metabolomics, lipidomics, metagenomics and systems biology.

Recommended Textbooks and References:
Course Objectives
The objectives of this course are to sensitize students about recent advances in molecular biology and various facets of molecular medicine which has potential to profoundly alter many aspects of modern medicine including pre- or post-natal analysis of genetic diseases and identification of individuals predisposed to disease ranging from common cold to cancer.

Student Learning Outcomes
Students should be able to understand various facets of molecular procedures and basics of genomics, proteomics and metabolomics that could be employed in early diagnosis and prognosis of human diseases.

Unit I
Genome biology in health and disease
4 lectures
DNA, RNA, Protein: An overview; chromosomal structure & mutations; DNA polymorphism: human identity; clinical variability and genetically determined adverse reactions to drugs.

Unit II
Genome: resolution, detection & analysis
5 lectures
PCR: Real-time; ARMS; Multiplex; ISH; FISH; ISA; RFLP; DHPLC; DGGE; CSCE; SSCP; Nucleic acid sequencing: new generations of automated sequencers; Microarray chips; EST; SAGE; microarray data normalization & analysis; molecular markers: 16S rRNA typing; Diagnostic proteomics: SELDI-TOF-MS; Bioinformatics data acquisition & analysis.

Unit III
Diagnostic metabolomics
2 lectures
Metabolite profile for biomarker detection the body fluids/tissues in various metabolic disorders by making using LCMS & NMR technological platforms.

Unit IV
Detection and identity of microbial diseases
4 lectures
Direct detection and identification of pathogenic-organisms that are slow growing or currently lacking a system of in vitro cultivation as well as genotypic markers of microbial resistance to specific antibiotics.

Unit V
Detection of inherited diseases
4 lectures
Exemplified by two inherited diseases for which molecular diagnosis has provided a dramatic improvement of quality of medical care: Fragile X Syndrome: Paradigm of new mutational mechanism of unstable triplet repeats, von-Hippel Lindau disease: recent acquisition in growing number of familial cancer syndromes.

Unit VI
Molecular oncology
5 lectures
Detection of recognized genetic aberrations in clinical samples from cancer patients; types of cancer-causing alterations revealed by next-generation sequencing of clinical isolates; predictive biomarkers for personalized onco-therapy of human diseases such as chronic myeloid leukemia, colon, breast, lung cancer and melanoma as well as matching targeted therapies with patients and preventing toxicity of standard systemic therapies.

Unit VII
Quality assurance and control
1 lecture
Quality oversight; regulations and approved testing.

Recommended Textbooks and References:
### Course Objectives

The objectives of this course are to give background on history of science, emphasizing methodologies used to do research, use framework of these methodologies for understanding effective lab practices and scientific communication and appreciate scientific ethics.

### Student Learning Outcomes

Students should be able to:
- Understand history and methodologies of scientific research, applying these to recent published papers;
- Understand and practice scientific reading, writing and presentations;
- Appreciate scientific ethics through case studies.

### Unit I

**History of science and science methodologies**

8 lectures

Empirical science; scientific method; manipulative experiments and controls; deductive and inductive reasoning; descriptive science; reductionist vs holistic biology.

### Unit II

**Preparation for research**

2 lectures

Choosing a mentor, lab and research question; maintaining a lab notebook.

### Unit III

**Process of communication**

5 lectures

Concept of effective communication- setting clear goals for communication; determining outcomes and results; initiating communication; avoiding breakdowns while communicating; creating value in conversation; barriers to effective communication; non-verbal communication-interpreting non-verbal cues; importance of body language, power of effective listening; recognizing cultural differences; Presentation skills - formal presentation skills; preparing and presenting using overhead projector, PowerPoint; defending interrogation; scientific poster preparation & presentation; participating in group discussions; Computing skills for scientific research - web browsing for information search; search engines and their mechanism of searching; hidden Web and its importance in scientific research; internet as a medium of interaction between scientists; effective email strategy using the right tone and conciseness.

### Unit IV

**Scientific communication**

9 lectures

Technical writing skills - types of reports; layout of a formal report; scientific writing skills - importance of communicating science; problems while writing a scientific document; plagiarism, software for plagiarism; scientific publication writing: elements of a scientific paper including abstract, introduction, materials & methods, results, discussion, references; drafting titles and framing abstracts; publishing scientific papers - peer review process and problems, recent developments such as open access and non-blind review; plagiarism; characteristics of effective technical communication; scientific presentations; ethical issues; scientific misconduct.
Recommended Textbooks and References:

Course Objectives
The objectives of this course are to provide students with experimental knowledge of molecular biology and genetic engineering.

Student Learning Outcomes
Students should be able to gain hands-on experience in gene cloning, protein expression and purification. This experience would enable them to begin a career in industry that engages in genetic engineering as well as in research laboratories conducting fundamental research.

Laboratory IV: Molecular Biology and Genetic Engineering

Syllabus

1. Concept of lac-operon:
   a) Lactose induction of B-galactosidase.
   b) Glucose Repression.
   c) Diauxic growth curve of E.coli
2. UV mutagenesis to isolate amino acid auxotroph
3. Phage titre with epsilon phage/M13
4. Genetic Transfer-Conjugation, gene mapping
5. Plasmid DNA isolation and DNA quantitation
6. Restriction Enzyme digestion of plasmid DNA
7. Agarose gel electrophoresis
8. Polymerase Chain Reaction and analysis by agarose gel electrophoresis
9. Vector and Insert Ligation
10. Preparation of competent cells
11. Transformation of E.coli with standard plasmids, Calculation of transformation efficiency
12. Confirmation of the insert by Colony PCR and Restriction mapping
13. Expression of recombinant protein, concept of soluble proteins and inclusion body formation in E.coli, SDS-PAGE analysis
14. Purification of His-Tagged protein on Ni-NTA columns
   a) Random Primer labeling
   b) Southern hybridization.

Recommended Textbooks and References:
Laboratory V: Immunology

Course Objectives
The objectives of this laboratory course are to develop an understanding about practical aspects of components of immune system as well as their function. Basic as well as advanced methods will be taught to detect different antigen and antibody interactions, isolation of different lymphocyte cells etc. and how they can be used in respective research work.

Student Learning Outcomes
Students should be able to:
• Evaluate usefulness of immunology in different pharmaceutical companies;
• Identify proper research lab working in area of their own interests;
• Apply their knowledge and design immunological experiments to demonstrate innate, humoral or cytotoxic T lymphocyte responses and figure out kind of immune responses in setting of infection (viral or bacterial) by looking at cytokine profile.

Syllabus
1. Selection of animals, preparation of antigens, immunization and methods of blood collection, serum separation and storage.
2. Antibody titre by ELISA method.
5. Isolation and purification of IgG from serum or IgY from chicken egg.
6. SDS-PAGE, Immunoblotting, Dot blot assays.
8. Separation of leucocytes by dextran method.
9. Demonstration of Phagocytosis of latex beads and their cryopreservation.
10. Separation of mononuclear cells by Ficoll-Hypaque and their cryopreservation.
11. Demonstration of ELISPOT.
12. Demonstration of FACS.

Semester Three
Bioprocess Engineering & Technology

Course Objectives
The objectives of this course are to educate students about the fundamental concepts of bioprocess technology and its related applications, thus preparing them to meet the challenges of the new and emerging areas of biotechnology industry.

Student Learning Outcomes
Students should be able to:
• Appreciate relevance of microorganisms from industrial context;
• Carry out stoichiometric calculations and specify models of their growth;
• Give an account of design and operations of various fermenters;
• Present unit operations together with the fundamental principles for basic methods in production technique for bio-based products;
• Calculate yield and production rates in a biological production process, and also interpret data;
• Calculate the need for oxygen and oxygen transfer;
• Critically analyze any bioprocess from market point of view;
• Give an account of important microbial/ enzymatic industrial processes in food and fuel industry.
<table>
<thead>
<tr>
<th>Unit I</th>
<th>Basic principles of biochemical engineering</th>
<th>4 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Isolation, screening and maintenance of industrially important microbes; microbial growth and death kinetics (an example from each group, particularly with reference to industrially useful microorganisms); strain improvement for increased yield and other desirable characteristics.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit II</th>
<th>Stoichiometry and models of microbial growth</th>
<th>4 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Elemental balance equations; metabolic coupling – ATP and NAD+; yield coefficients; unstructured models of microbial growth; structured models of microbial growth.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit III</th>
<th>Bioreactor design and analysis</th>
<th>8 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Batch and continuous fermenters; modifying batch and continuous reactors: chemostat with recycle, multistage chemostat systems, fed-batch operations; conventional fermentation v/s biotransformation; immobilized cell systems; large scale animal and plant cell cultivation; fermentation economics; upstream processing: media formulation and optimization; sterilization; aeration, agitation and heat transfer in bioprocess; scale up and scale down; measurement and control of bioprocess parameters.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit IV</th>
<th>Downstream processing and product recovery</th>
<th>8 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Separation of insoluble products - filtration, centrifugation, sedimentation, flocculation; Cell disruption; separation of soluble products: liquid-liquid extraction, precipitation, chromatographic techniques, reverse osmosis, ultra and micro filtration, electrophoresis; final purification: drying; crystallization; storage and packaging.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit V</th>
<th>Fermentation economics</th>
<th>4 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Isolation of micro-organisms of potential industrial interest; strain improvement; market analysis; equipment and plant costs; media; sterilization, heating and cooling; aeration and agitation; bath-process cycle times and continuous cultures; recovery costs; water usage and recycling; effluent treatment and disposal.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit VI</th>
<th>Applications of enzyme technology in food processing</th>
<th>4 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mechanism of enzyme function and reactions in process techniques; enzymatic bioconversions e.g. starch and sugar conversion processes; high-fructose corn syrup; interesterified fat; hydrolyzed protein etc. and their downstream processing; baking by amylases, deoxygenation and desugaring by glucose oxidase, beer mashing and chill proofing; cheese making by proteases and various other enzyme catalytic actions in food processing.</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit VII</th>
<th>Applications of microbial technology in food process operations and production, biofuels and biorefinery</th>
<th>4 lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Fermented foods and beverages; food ingredients and additives prepared by fermentation and their purification; fermentation as a method of preparing and preserving foods; microbes and their use in pickling, producing colours and flavours, alcoholic beverages and other products; process wastes-whey, molasses, starch substrates and other food wastes for bioconversion to useful products; bacteriocins from lactic acid bacteria – production and applications in food preservation; biofuels and biorefinery</td>
<td></td>
</tr>
</tbody>
</table>

**Recommended Textbooks and References:**

Course Objectives

This course is broad-based in nature encompassing several new technologies that current experimental researchers are employing to probe complex system biology questions in life-sciences. The objectives of this course are to teach basics of the new principles to students so as to appreciate current-day research tool-kit better.

Student Learning Outcomes

Students should be able to learn the history, theoretical basis and basic understanding of these technologies in area of biotechnology. They should also be able to learn about various applications of these technologies. The students may also learn one application in depth through an assignment and/or seminar.

Emerging Technologies

Credits

2

Unit I
Optical microscopy methods
8 lectures

Basic Microscopy: Light Microscopy: lenses and microscopes, resolution: Rayleigh’s Approach, Darkfield; Phase Contrast; Differential Interference Contrast; fluorescence and fluorescence microscopy: what is fluorescence, what makes a molecule fluorescent, fluorescence microscope; optical arrangement, light source; filter sets: excitation filter, dichroic mirror, and barrier, optical layout for image capture; CCD cameras; back illumination, binning; recording color; three CCD elements with dichroic beamsplitters, boosting the signal.

Advanced Microscopy: Confocal microscope: scanning optical microscope, confocal principle, resolution and point spread function, light source: gas lasers & solid-state, primary beamsplitter; beam scanning, pinhole and signal channel configurations, detectors; pixels and voxels; contrast, spatial sampling: temporal sampling: signal-to-noise ratio, multichannel images. nonlinear microscopy; multiphoton microscopy; principles of two-photon fluorescence, advantages of two-photon excitation, tandem scanning (spinning disk) microscopes, deconvolving confocal images; image processing, three-dimensional reconstruction; advanced fluorescence techniques: FLIM, FRET, and FCS, Fluorescence Lifetime, Fluorescence Resonant Energy Transfer (FRET), Fluorescence Correlation Spectroscopy (FCS), Evanescent Wave Microscopy; Near-Field and Evanescent Waves, Total Internal Reflection Microscopy; Near-Field Microscopy; Beyond the Diffraction Limit: Stimulated Emission Depletion (STED), Super-Resolution Summary, Super-Resolution Imaging with Stochastic Optical Reconstruction Microscopy (STORM) and Photoactivated Localization Microscopy (PALM).

Unit II
Mass spectroscopy
4 lectures

Ionization techniques; mass analyzers/overview MS; FT-ICR and Orbitrap, fragmentation of peptides; proteomics, nano LC-MS; Phospho proteomics; interaction proteomics, mass spectroscopy in structural biology; imaging mass spectrometry.

Unit III
Systems biology
3 lectures

High throughput screens in cellular systems, target identification, validation of experimental methods to generate the omics data, bioinformatics analyses, mathematical modeling and designing testable predictions.

Unit IV
Structural biology
3 lectures


Unit V
CRISPR-CAS
6 lectures

History of its discovery, elucidation of the mechanism including introduction to all the molecular players, development of applications for in vivo genome engineering for genetic studies, promise of the technology as a next generation therapeutic method.
Introduction to nanobodies, combining nanobody with phage-display method for development of antibody against native proteins, nanobody as a tool for protein structure-function studies, use of nanobodies for molecular imaging, catabolic antibodies using nanobodies.

Recommended Textbooks and References:
Note: This paper demonstrates that DNA is the transforming Principle originally described by Fredrick Griffith.

Note: This paper demonstrates that DNA, and not protein, component of phages enter bacterial cells.

3. Molecular structure of nucleic acids; a structure for deoxyribose nucleic acid Watson JD and Crick FH; Nature. 1953 Apr 25;171(4356):737-8 
Note: In this one page paper Watson and Crick first described the structure of DNA double helix

4. Transposable mating type genes in Saccharomyces cerevisiae James Hicks, Jeffrey N. Strathern & Amar J.S. Klar; Nature 282, 478-483, 1979 
Note: This paper provided evidence for 'cassette hypothesis' of yeast mating type switches i.e. interconversion of mating types in yeast (S. cerevisiae) occurs by DNA rearrangement.

Note: The experiment demonstrating semi-conservative mode of DNA replication is referred to as "the most beautiful experiment in biology"

Note: This paper demonstrates that the telomerase contains the template for telomere synthesis

---

How does the Course Module work? Students may be divided in groups and each group may be responsible for one classical paper. Each week there may be a 1.5 hour presentation cum discussion for each of the papers. At the end of the semester each student will be asked to write a mini-review (2-3 pages long) on any one classical paper, other than the one he/she presented/discussed.

A list of sixteen classic papers and some suggested reference materials:

### Syllabus

#### Molecular Biology

   Note: This paper demonstrates that DNA is the transforming Principle originally described by Fredrick Griffith.

   Note: Note: This paper demonstrates that DNA, and not protein, component of phages enter bacterial cells.

3. Molecular structure of nucleic acids; a structure for deoxyribose nucleic acid Watson JD and Crick FH; Nature. 1953 Apr 25;171(4356):737-8 
   Note: In this one page paper Watson and Crick first described the structure of DNA double helix

4. Transposable mating type genes in Saccharomyces cerevisiae James Hicks, Jeffrey N. Strathern & Amar J.S. Klar; Nature 282, 478-483,1979 
   Note: This paper provided evidence for 'cassette hypothesis' of yeast mating type switches i.e. interconversion of mating types in yeast (S. cerevisiae) occurs by DNA rearrangement.

   Note: The experiment demonstrating semi-conservative mode of DNA replication is referred to as "the most beautiful experiment in biology"

   Note: This paper demonstrates that the telomerase contains the template for telomere synthesis

---

#### Cell Biology

   Note: This paper demonstrates the existence of a protein conducting channel

Study help - A brief history of Signal Hypothesis
2. Identification of 23 complementation groups required for post-translational events in the yeast secretory pathway
Note: In this groundbreaking paper Randy Schekman’s group used a mutagenesis screen for fast sedimenting yeast mutants to identify genes involved in cell secretion

3. A yeast mutant defective at an early stage in import of secretory protein precursors into the endoplasmic reticulum
Note: Using another yeast mutation screen Schekman lab identifies Sec61, a component of ER protein Conducting Channel (PCC)
Suggested reference paper - A biochemical assay for identification of PCC.

4. Reconstitution of the Transport of Protein between Successive Compartments of the Golgi
Balch WE, Dunphy WG, Braell WA, Rothman JE.; Cell. 1984 Dec;39(2 Pt 1):405-16
Note: This paper describes setting up of an in vitro reconstituted system for transport between golgi stacks which eventually paved the way for identification of most of the molecular players involved in these steps including NSF, SNAP etc.

5. A complete immunoglobulin gene is created by somatic recombination
Note: This study demonstrates DNA level molecular details of somatic rearrangement of immunoglobulin gene sequences leading to the generation of functionally competent antibody generating gene following recombination.

6. A novel multigene family may encode odorant receptors: a molecular basis for odor recognition
Buck I and Axel R; Cell. 1991 Apr 5;65(1):175-87
Note: This paper suggests that different chemical odorants associate with different cell-specific expression of a transmembrane receptor in Drosophila olfactory epithelium where a large family of odorat receptors is expressed.

7. Kinesin walks hand-over-hand
Yildiz A, Tomishige M, Vale RD, Selvin PR.; Science. 2004 Jan 30;303(5658):676-8
Note: This paper shows that kinesin motor works as a two-headed dimeric motor walking hand-over-hand rather than like an inchworm on microtubule tract using the energy of ATP hydrolysis.

Syllabus
Developmental Biology/ Genetics

1. Mutations affecting segment number and polarity in Drosophila
Note: This single mutagenesis screen identified majority of the developmentally important genes not only in flies but in other metazoans as well.

2. Information for the dorsal--ventral pattern of the Drosophila embryo is stored as maternal mRNA
Anderson KV and Nüsslein-Volhard C; Nature. 1984 Sep 20-26;311(5983):223-7
Note: This landmark paper demonstrated that early dorsal-ventral pattern information is stored as maternal mRNA in flies and devised the method of identifying genes encoding such genes.

3. Hedgehog signalling in the mouse requires intrflagellar transport proteins
Note: One of the architects of original fly mutagenesis screens conducted a mouse mutagenes screen which identified a gene Kif5a as a major component of hedgehog signaling pathway. Eventually this discovery revolutionizes our understanding of mechanisms of action of signaling pathways by demonstrating central role of cilia in it.
Suggested Reference paper - Design and execution of a embryonic lethal mutation screen in mouse.
Bioentrepreneurship

Credits

2

Course Objectives

Research and business belong together and both are needed. In a rapidly developing life science industry, there is an urgent need for people who combine business knowledge with the understanding of science & technology. Bio-entrepreneurship, an interdisciplinary course, revolves around the central theme of how to manage and develop life science companies and projects. The objectives of this course are to teach students about concepts of entrepreneurship including identifying a winning business opportunity, gathering funding and launching a business, growing and nurturing the organization and harvesting the rewards.

Unit I
Innovation and entrepreneurship in bio-business
8 lectures

Introduction and scope in Bio-entrepreneurship, Types of bio-industries and competitive dynamics between the sub-industries of the bio-sector (e.g. pharmaceuticals vs. Industrial biotech), Strategy and operations of bio-sector firms: Factors shaping opportunities for innovation and entrepreneurship in bio-sectors, and the business implications of those opportunities, Alternatives faced by emerging bio-firms and the relevant tools for strategic decision, Entrepreneurship development programs of public and private agencies (MSME, DBT, BIRAC, Make In India), strategic dimensions of patenting & commercialization strategies.

Unit II
Bio markets - business strategy and marketing
8 lectures

Negotiating the road from lab to the market (strategies and processes of negotiation with financiers, government and regulatory authorities), Pricing strategy, Challenges in marketing in bio business (market conditions & segments; developing distribution channels, the nature, analysis and management of customer needs), Basic contract principles, different types of agreement and contract terms typically found in joint venture and development agreements, Dispute resolution skills.

Unit III
Finance and accounting
8 lectures

Business plan preparation including statutory and legal requirements, Business feasibility study, financial management issues of procurement of capital and management of costs, Collaborations & partnership, Information technology.

Unit IV
Technology management
8 lectures

Technology – assessment, development & upgradation, Managing technology transfer, Quality control & transfer of foreign technologies, Knowledge centers and Technology transfer agencies, Understanding of regulatory compliances and procedures (CDSCO, NBA, GCP, GLA, GMP).

Recommended Textbooks and References:


---

**Course Objectives**

The objectives of this course are:

- To provide basic knowledge on intellectual property rights and their implications in biological research and product development;
- To become familiar with India’s IPR Policy;
- To learn biosafety and risk assessment of products derived from biotechnology and regulation of such products;
- To become familiar with ethical issues in biological research. This course will focus on consequences of biomedical research technologies such as cloning of whole organisms, genetic modifications, DNA testing.

**Student Learning Outcomes**

On completion of this course, students should be able to:

- Understand the rationale for and against IPR and especially patents;
- Understand why India has adopted an IPR Policy and be familiar with broad outline of patent regulations;
- Understand different types of intellectual property rights in general and protection of products derived from biotechnology research and issues related to application and obtaining patents;
- Gain knowledge of biosafety and risk assessment of products derived from recombinant DNA research and environmental release of genetically modified organisms, national and international regulations;
- Understand ethical aspects related to biological, biomedical, health care and biotechnology research.

**Unit I**

**Introduction to IPR**

- 5 lectures

Introduction to intellectual property; types of IP: patents, trademarks, copyright & related rights, industrial design, traditional knowledge, geographical indications, protection of new GMOs; International framework for the protection of IP; IP as a factor in R&D; IPs of relevance to biotechnology and few case studies; introduction to history of GATT, WTO, WIPO and TRIPS; plant variety protection and farmers rights act; concept of ‘prior art’: invention in context of “prior art”; patent databases - country-wise patent searches (USPTO, EPO, India); analysis and report formation.

**Unit II**

**Patenting**

- 5 lectures

Basics of patents: types of patents; Indian Patent Act 1970; recent amendments; WIPO Treaties; Budapest Treaty; Patent Cooperation Treaty (PCT) and implications; procedure for filing a PCT application; role of a Country Patent Office; filing of a patent application; precautions before patenting-disclosure/non-disclosure - patent application- forms and guidelines including those of National Bio-diversity Authority (NBA) and other regulatory bodies, fee structure, time frames; types of patent applications: provisional and complete specifications; PCT and conventional patent applications; international patenting-requirement, procedures and costs; financial assistance for patenting-introduction to existing schemes; publication of patents-gazette of India, status in Europe and US; patent infringement- meaning, scope, litigation, case studies and examples; commercialization of patented innovations; licensing – outright sale, licensing, royalty; patenting by research students and scientists-university/organizational rules in India and abroad, collaborative research - backward and forward IP; benefit/credit sharing among parties/community, commercial (financial) and non-commercial incentives.
Unit III
Biosafety
5 lectures
Biosafety and Biosecurity - introduction; historical background; introduction to biological safety cabinets; primary containment for biohazards; biosafety levels; GRAS organisms, biosafety levels of specific microorganisms; recommended biosafety levels for infectious agents and infected animals; definition of GMOs & LMOs; principles of safety assessment of transgenic plants – sequential steps in risk assessment; concepts of familiarity and substantial equivalence; risk – environmental risk assessment and food and feed safety assessment; problem formulation – protection goals, compilation of relevant information, risk characterization and development of analysis plan; risk assessment of transgenic crops vs cisgenic plants or products derived from RNAi, genome editing tools.

Unit IV
National and international regulations
5 lectures
International regulations – Cartagena protocol, OECD consensus documents and Codex Alimentarius; Indian regulations – EPA act and rules, guidance documents, regulatory framework – RCGM, GEAC, IBSC and other regulatory bodies; Draft bill of Biotechnology Regulatory authority of India - containments – biosafety levels and category of rDNA experiments; field trails – biosafety research trials – standard operating procedures - guidelines of state governments; GM labeling – Food Safety and Standards Authority of India (FSSAI).

Unit V
Bioethics
5 lectures

Recommended Textbooks and References:
2. National IPR Policy, Department of Industrial Policy & Promotion, Ministry of Commerce, GoI
5. Office of the Controller General of Patents, Design & Trademarks; Department of Industrial Policy & Promotion; Ministry of Commerce & Industry; Government of India. http://www.ipindia.nic.in/


Course Objectives
The purpose of this course is to help students organize ideas, material and objectives for their dissertation and to begin development of communication skills and to prepare the students to present their topic of research and explain its importance to their fellow classmates and teachers.

Student Learning Outcomes
Students should be able to demonstrate the following abilities:

- Formulate a scientific question;
- Present scientific approach to solve the problem;
- Interpret, discuss and communicate scientific results in written form;
- Gain experience in writing a scientific proposal;
- Learn how to present and explain their research findings to the audience effectively.

Syllabus
Project Proposal Preparation

Selection of research lab and research topic: Students should first select a lab wherein they would like to pursue their dissertation. The supervisor or senior researchers should be able to help the students to read papers in the areas of interest of the lab and help them select a topic for their project. The topic of the research should be hypothesis driven.

Review of literature: Students should engage in systematic and critical review of appropriate and relevant information sources and appropriately apply qualitative and/or quantitative evaluation processes to original data; keeping in mind ethical standards of conduct in the collection and evaluation of data and other resources.

Writing Research Proposal: With the help of the senior researchers, students should be able to discuss the research questions, goals, approach, methodology, data collection, etc. Students should be able to construct a logical outline for the project including analysis steps and expected outcomes and prepare a complete proposal in scientific proposal format for dissertation.

Syllabus
Poster Presentation

Students will have to present the topic of their project proposal after few months of their selection of the topic. They should be able to explain the novelty and importance of their research topic.

Syllabus
Oral Presentation

At the end of their project, presentation will have to be given by the students to explain work done by them in detail. Along with summarizing their findings they should also be able to discuss the future expected outcome of their work.
Laboratory VI: Bioprocess Engineering & Technology

Course Objectives
The objectives of this laboratory course are to provide hands-on training to students in upstream and downstream unit operations.

Student Learning Outcomes
Students should be able to:
• Investigate, design and conduct experiments, analyze and interpret data, and apply the laboratory skills to solve complex bioprocess engineering problems;
• Apply skills and knowledge gained will be useful in solving problems typical of bio industries and research.

Credits 4

Syllabus

1. Basic Microbiology techniques
   a) Scale up from frozen vial to agar plate to shake flask culture.
   b) Instrumentation: Microplate reader, spectrophotometer, microscopy.
   c) Isolation of microorganisms from soil samples.

2. Experimental set-up
   a) Assembly of bioreactor and sterilization.
   b) Growth kinetics.
   c) Substrate and product inhibitions.
   d) Measurement of residual substrates.

3. Data Analysis
   a) Introduction to Metabolic Flux Analysis (MFA).

4. Fermentation
   a) Batch.
   b) Fed-batch.
   c) Continuous.

5. Unit operations
   a) Microfiltrations: Separation of cells from broth.
   b) Bioseparations: Various chromatographic techniques and extractions.

6. Bioanalytics
   a) Analytical techniques like HPLC, FPLC, GC, GC-MS etc. for measurement of amounts of products/substrates.

Recommended Textbooks and References:
Course Objectives
The aim of this course is to provide practical training in bioinformatic methods including accessing major public sequence databases, use of different computational tools to find sequences, analysis of protein and nucleic acid sequences by various software packages.

Student Learning Outcomes
On completion of this course, students should be able to:

• Describe contents and properties of most important bioinformatics databases;
• Perform text- and sequence-based searches and analyze and discuss results in light of molecular biological knowledge;
• Explain major steps in pairwise and multiple sequence alignment, explain principle and execute pairwise sequence alignment by dynamic programming;
• Predict secondary and tertiary structures of protein sequences.

Syllabus

1. Using NCBI and Uniprot web resources.
2. Introduction and use of various genome databases.
4. Similarity searches using tools like BLAST and interpretation of results.
5. Multiple sequence alignment using ClustalW.
7. Use of gene prediction methods (GRAIL, Genscan, Glimmer).
8. Using RNA structure prediction tools.
9. Use of various primer designing and restriction site prediction tools.
10. Use of different protein structure prediction databases (PDB, SCOP, CATH).
11. Construction and study of protein structures using Deepview/PyMol.
13. Use of tools for mutation and analysis of the energy minimization of protein structures.
14. Use of miRNA prediction, designing and target prediction tools.

Semester Four

Dissertation

Course Objectives
The objectives of this course are to prepare the students to adapt to the research environment and understand how projects are executed in a research laboratory. It will also enable students to learn practical aspects of research and train students in the art of analysis and thesis writing.

Student Learning Outcomes
Students should be able to learn how to select and defend a topic of their research, how to effectively plan, execute, evaluate and discuss their experiments. Students should be able to demonstrate considerable improvement in the following areas:

• In-depth knowledge of the chosen area of research.
• Capability to critically and systematically integrate knowledge to identify issues that must be addressed within framework of specific thesis.
• Competence in research design.
Recommended Electives

**Biological Imaging**

| Credits | 2 |

**Course Objectives**
The objectives of this course are to provide complete overview of state-of-art live-cell imaging techniques using microscopes currently available in literature. Live-cell imaging techniques allow real-time examination of almost every aspect of cellular function under normal and experimental conditions. With live-cell imaging experiments, main challenges are to keep cells alive and healthy over a period of time. The growing number of live-cell imaging techniques means one can obtain greater amounts of information without stressing out cells.

**Student Learning Outcomes**
On completion of this course, students shall be able to gain a complete overview of super-resolution field from fundamentals to state-of-art methods and applications in biomedical research. The students shall learn the comparative advantages and disadvantages of each technique, covers all key techniques in field of biomedical science. The students shall also learn how to use new tools to increase resolution in sub-nanometer-scale images of living cells and tissue, which leads to new information about molecules, pathways and dynamics and state-of-the-art examples of applications using microscopes.

**Unit I**

**Widefield fluorescent microscopy**

3 lectures

One of the most basic techniques for live-cell imaging is widefield fluorescent microscopy. Standard inverted research grade microscopes can yield valuable results if you are imaging adherent cells, large regions of interest (such as organelles) or very thin tissue sections (less than 5 micrometer). In widefield, a CCD camera is usually used to...
capture images and the epi-fluorescence illumination source can be a mercury lamp, xenon lamp, LED's, etc. Each of light sources require carefully matched interference filters for specific excitation and emission wavelengths of your fluorophore of interest. With widefield microscopy, your specimen is only exposed to excitation light for relatively short time periods as the full aperture of emission light is collected by the objectives. Widefield fluorescence microscopy can be used in combination with other common contrast techniques such as phase contrast and differential interference contrast (DIC) microscopy. This combination is useful when performing live-cell imaging to examine general cell morphology or viability while also imaging regions of interest within cells.

Unit II
Confocal laser scanning microscopy (CLSM)
3 lectures
CLSM has ability to eliminate out-of-focus light and information. It is also possible to obtain optical serial sections from thicker specimens. A conjugate pinhole in optical path of confocal microscope prevents fluorescence from outside of focal plane from being collected by photomultiplier detector or imaged by camera. In CLSM, a single pinhole (and single focused laser spot) is scanned across specimen by scanning system. This spot forms a reflected epi-fluorescence image back on original pinhole. When specimen is in focus, fluorescent light from it passes through pinhole to detector. Any out-of-focus light is defocused at pinhole and very little of this signal passes through to detector meaning that background fluorescence is greatly reduced. The pinhole acts as a spatial filter for emission light from the specimen.

Unit III
Spinning disc confocal microscopy (SDCM)
2 lectures
This method utilises a 'Nipkow Disc' which is a mechanical opaque disc which has a series of thousands of drilled or etched pinholes arranged in a spiral pattern. Each illuminated pinhole on disc is imaged by microscope objective to a diffraction-limited spot on region of interest on specimen. The emission from fluorophores passes back through Nipkow disc pinholes and can be observed and captured by a CCD camera. The effect of spinning disc is that many thousands of points on specimen are simultaneously illuminated. Using SDCM to examine a specimen means that real-time imaging (30-frames-per-second or faster) can be achieved, which is extremely useful if you are looking at dynamic changes within living cells over a wide spectrum of time-scales.

Unit IV
Light-sheet fluorescence microscopy (LSFM, or SPIM)
2 lectures
This method enables one to perform live-cell imaging on whole embryos, tissues and cell spheroids in vivo in a gentle manner with high temporal resolution and in three dimensions. One is able to track cell movement over extended periods of time and follow development of organs and tissues on a cellular level. The next evolution of light-sheet fluorescence microscopy, termed lattice light-sheet microscopy as developed by Eric Betzig (Nobel Prize Laureate 2014 for PALM super-resolution microscopy) will even allow live-cell imaging with super-resolved in vivo cellular localization capabilities.

Unit V
Super-resolved fluorescence microscopy
8 lectures

Unit VI
Re-scan confocal microscopy
4 lectures
Structured Illumination Microscopy; Correlative Nanoscopy: AFM Super-Resolution (STED/STORM) ; Stochastic Optical Fluctuation Imaging.
Course Objectives
The objective of this course is to provide students with theory and practical experience of essentials to aid for genomic, proteomic and metabolomics courses and drug design program.

Student Learning Outcomes
On completion of this course, the students are expected to:
• Develop an understanding of the basic theory of these computational tools;
• Develop required database extraction, integration, coding for computational tools and methods necessary for all Omics;
• Create hypothesis for investigating specific contemporary biological questions, provide help to experiment with or develop appropriate tools;
• Critically analyze and interpret results of their study with respect to whole systems.

Unit I
Introduction to computational biology basics and biological databases
4 lectures

Computers in biology and medicine; Overview of biological databases, nucleic acid & protein databases, primary, secondary, functional, composite, structural classification database, Sequence formats & storage. Access databases, Extract and create sub databases, limitations of existing databases.

Unit II
Pairwise and multiple sequence alignments
5 lectures


Unit III
Genome analysis
6 lectures

Polymorphisms in DNA sequence, Introduction to Next Generation Sequencing technologies, Whole Genome Assembly and challenges, Sequencing and analysis of large genomes, Gene prediction, Functional annotation, Comparative genomics, Probabilistic functional gene networks, Human genome project, Genomics and crop improvement. Study available GWAS, ENCODE, HUGO projects, extract and build sub databases; Visualization tools including Artemis and Vista for genome comparison; Functional genomics case studies.

Unit IV
Structure visualization
3 lectures

Retrieving and drawing structures, Macromolecule viewing platforms, Structure validation and correction, Structure optimization, Analysis of ligand-protein interactions; Tools such as PyMol or VMD.
This course will give a broad overview of research and development carried out in industrial setup towards drug discovery.

**Student Learning Outcomes**

On completion of this course, students should be able to understand basics of R&D in drug discovery and should be able to apply knowledge gained in respective fields of pharmaceutical industry.

**Recommended Textbooks and References:**


**Course Objectives**

This course will give a broad overview of research and development carried out in industrial setup towards drug discovery.

**Unit I**

**Target identification and molecular modelling**

7 lectures

Identification of target or drug leads associated with a particular disease by a number of different techniques including combinations of molecular modeling, combinatorial libraries and high-throughput screening (HTS); Conceptualizing the automation of the HTS process and the importance of bioinformatics and data processing in identification of lead compounds; Rational drug design, based on understanding the three-dimensional

**Unit V**

**Molecular modelling**

6 lectures

Significance and need, force field methods, energy, buried and exposed residues; side chains and neighbours; fixed regions; hydrogen bonds; mapping properties onto surfaces; RMS fit of conformers and protein chains, assigning secondary structures; sequence alignment: methods, evaluation, scoring; protein curation: backbone construction and side chain addition; different types of protein chain modelling: ab initio, homology, hybrid, loop; Template recognition and alignments; Modelling parameters and considerations; Model analysis and validation; Model optimization; Substructure manipulations, annealing, protein folding and model generation; loop generating methods; loop analysis; Analysis of active sites using different methods in studying protein–protein interactions.

**Unit VI**

**Structure-based drug development**

6 lectures

Molecular docking: Types and principles, Semi-flexible docking, Flexible docking; Ligand and protein preparation, Macromolecule and ligand optimization, Ligand conformations, Clustering, Analysis of docking results and validation with known information. Extra-precision docking platforms, Use of Small-molecule libraries, Natural compound libraries for virtual high throughput screenings.

**Unit VII**

**Ligand-based drug development**

6 lectures

Quantitative structure activity relationships; Introduction to chemical descriptors like 2D, 3D and Group-based; Radar plots and contribution plots and Activity predictions, Pharmacophore modeling, Pharmacophore-based screenings of compound library, analysis and experimental validation.
structures and physicochemical properties of drugs and receptors; Modelling drug/receptor interactions with the emphasis on molecular mechanisms, molecular dynamics simulations and homology modelling; Conformational sampling, macromolecular folding, structural bioinformatics, receptor-based and ligand-based design and docking methods, in silico screening of libraries, semi-empirical and ab-initio methods, QSAR methods, molecular diversity, design of combinatorial libraries of drug-like molecules, macromolecular and chemical databases.

Unit II
Lead optimization
5 lectures
Identification of relevant groups on a molecule that interact with a receptor and are responsible for biological activity; Understanding structure activity relationship; Structure modification to increase potency and therapeutic index; Concept of quantitative drug design using Quantitative structure–activity relationship models (QSAR models) based on the fact that the biological properties of a compound are a function of its physicochemical parameters such as solubility, lipophilicity, electronic effects, ionization, stereochemistry, etc.; Bioanalytical assay development in support of in vitro and in vivo studies (LC/MS/MS, GC/MS and ELISA).

Unit III
Preclinical development
5 lectures
Principles of drug absorption, drug metabolism and distribution - intestinal absorption, metabolic stability, drug-drug interactions, plasma protein binding assays, metabolite profile studies, Principles of toxicology, Experimental design for preclinical and clinical PK/PD/TK studies, Selection of animal model; Regulatory guidelines for preclinical PK/PD/TK studies; Scope of GLP, SOP for conduct of clinical & non clinical testing, control on animal house, report preparation and documentation Integration of non-clinical and preclinical data to aid design of clinical studies.

Unit IV
Drug manufacturing
4 lectures
Requirements of GMP implementation, Documentation of GMP practices, CoA, Regulatory certification of GMP, Quality control and Quality assurance, concept and philosophy of TQM, ICH and ISO 9000; ICH guidelines for Manufacturing, Understanding Impurity Qualification Data, Stability Studies.

Unit V
Clinical trial design
4 lectures
Objectives of Phase I, II, III and IV clinical studies, Clinical study design, enrollment, sites and documentation, Clinical safety studies: Adverse events and adverse drug reactions, Clinical PK, pharmacology, drug-drug interaction studies, Statistical analysis and documentation.

Unit VI
Fundamentals of regulatory affairs and bioethics
4 lectures
Global Regulatory Affairs and different steps involved, Regulatory Objectives, Regulatory Agencies; FDA guidelines on IND and NDA submissions, Studies required for IND and NDA submissions for oncology, HIV, cardiovascular indications, On-label vs. off-label drug use GCP and Requirements of GCP Compliance, Ethical issues and Compliance to current ethical guidelines, Ethical Committees and their set up, Animal Ethical issues and compliance.

Recommended Textbooks and References:
**Course Objectives**
This course aims to introduce fundamentals of Environmental Biotechnology. The course will introduce major groups of microorganisms—tools in biotechnology and their most important environmental applications. The environmental applications of biotechnology will be presented in detail and will be supported by examples from the national and international literature.

**Student Learning Outcomes**
On completion of course, students will be able to understand use of basic microbiological, molecular and analytical methods, which are extensively used in environmental biotechnology.

**Unit I**
**Introduction to Environment**
6 lectures

Introduction to environment; pollution and its control; pollution indicators; waste management: domestic, industrial, solid and hazardous wastes; strain improvement; Biodiversity and its conservation; Role of microorganisms in geochemical cycles; microbial energy metabolism, microbial growth kinetics and elementary chemostat theory, relevant microbiological processes, microbial ecology.

**Unit II**
**Bioremediation**
6 lectures

Bioremediation: Fundamentals, methods and strategies of application (biostimulation, bioaugmentation) – examples, bioremediation of metals (Cr, As, Se, Hg), radionuclides (U, Te), organic pollutants (PAHs, PCBs, Pesticides, TNT etc.), technological aspects of bioremediation (in situ, ex situ).

**Unit III**
**Role of microorganisms in bioremediation**
6 lectures

Application of bacteria and fungi in bioremediation: White rot fungi vs specialized degrading bacteria: examples, uses and advantages vs disadvantages; Phytoremediation: Fundamentals and description of major methods of application (phytoaccumulation, phytovolatilization, rhizofiltration phytostabilization).

**Unit IV**
**Biotechnology and agriculture**
11 lectures

Bioinsecticides: *Bacillus thuringiensis*, Baculoviruses, uses, genetic modifications and aspects of safety in their use; Biofungicides: Description of mode of actions and mechanisms (e.g. *Trichoderma, Pseudomonas fluorescens*); Biofertilizers: Symbiotic systems between plants – microorganisms (nitrogen fixing symbiosis, mycorrhiza fungi symbiosis), Plant growth promoting rhizobacteria (PGPR) – uses, practical aspects and problems in application.

**Unit V**
**Biofuels**
11 lectures

Environmental Biotechnology and biofuels: biogas; bioethanol; biodiesel; biohydrogen; Description of the industrial processes involved, microorganisms and biotechnological interventions for optimization of production; Microbiologically enhanced oil recovery (MEOR); Biodeleaching of metals; Production of bioplastics; Production of biosurfactants: bioemulsifiers; Paper production: use of xylanases and white rot fungi.

**Recommended Textbooks and References:**

**Microbial Technology**

**Course Objectives**
The objectives of this course are to introduce students to developments/advances made in field of microbial technology for use in human welfare and solving problems of the society.

**Student Learning Outcomes**
On completion of this course, students would develop deeper understanding of the microbial technology and its applications.

<table>
<thead>
<tr>
<th>Unit</th>
<th>Introduction to microbial technology</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>8 lectures</td>
</tr>
<tr>
<td></td>
<td><strong>Microbial technology in human welfare; Isolation and screening of microbes important for industry – advances in methodology and its application; Advanced genome and epigenome editing tools (e.g., engineered zinc finger proteins, TALEs/TALENs, and the CRISPR/Cas9 system as nucleases for genome editing, transcription factors for epigenome editing, and other emerging tools) for manipulation of useful microbes/strains and their applications; Strain improvement to increase yield of selected molecules, e.g., antibiotics, enzymes, biofuels.</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit</th>
<th>Environmental applications of microbial technology</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>6 lectures</td>
</tr>
<tr>
<td></td>
<td><strong>Environmental application of microbes; Ore leaching; Biodegradation - biomass recycle and removal; Bioremediation - toxic waste removal and soil remediation; Global Biogeochemical cycles; Environment sensing (sensor organisms/biological sensors); International and National guidelines regarding use of genetically modified organisms in environment, food and pharmaceuticals.</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit</th>
<th>Pharmaceutical applications of microbial technology</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>8 lectures</td>
</tr>
<tr>
<td></td>
<td><strong>Recombinant protein and pharmaceuticals production in microbes – common bottlenecks and issues (technical/operational, commercial and ethical); Attributes required in industrial microbes (Streptomyces sp., Yeast) to be used as efficient cloning and expression hosts (biologicals production); Generating diversity and introduction of desirable properties in industrially important microbes (Streptomyces/Yeast); Microbial cell factories; Downstream processing approaches used in industrial production process (Streptomyces sp., Yeast).</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit</th>
<th>Food applications of microbial technology</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>7 lectures</td>
</tr>
<tr>
<td></td>
<td><strong>Application of microbes and microbial processes in food and healthcare industries - food processing and food preservation, antibiotics and enzymes production, microbes in targeted delivery application – drugs and vaccines (bacterial and viral vectors); Non-recombinant ways of introducing desirable properties in Generally recognized as safe (GRAS) microbes to be used in food (e.g., Yeast) - exploiting the existing natural diversity or the artificially introduced diversity through conventional acceptable techniques (mutagenesis, protoplast fusion, breeding, genome shuffling, directed evolution etc.).</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Unit</th>
<th>Advances in microbial technology</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>8 lectures</td>
</tr>
<tr>
<td></td>
<td><strong>Microbial genomics for discovery of novel enzymes, drugs/antibiotics; Limits of microbial genomics with respect to use in human welfare; Metagenomics and metatranscriptomics – their potential, methods to study and applications/use (animal and plant health, environmental clean-up, global nutrient cycles &amp; global sustainability, understanding evolution), Global metagenomics initiative - surveys/projects and outcome, metagenomic library construction and functional screening in suitable hosts – tools and techniques for discovery/identification of novel enzymes, drugs (e.g., protease, antibiotic) etc.</strong></td>
</tr>
</tbody>
</table>

| Credits | 2 |

<table>
<thead>
<tr>
<th>2 Unit I Introduction to microbial technology 8 lectures</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>6 Unit II Environmental applications of microbial technology 6 lectures</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>8 Unit III Pharmaceutical applications of microbial technology 8 lectures</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>7 Unit IV Food applications of microbial technology 7 lectures</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>8 Unit V Advances in microbial technology 8 lectures</th>
</tr>
</thead>
</table>
Course Objectives
The aim of this course is to introduce methods and strategies commonly used in protein engineering.

Student Learning Outcomes
On completion of this course, students should be able to:

- Analyse structure and construction of proteins by computer-based methods;
- Describe structure and classification of proteins;
- Analyse purity and stability of proteins and explain how to store them in best way;
- Explain how proteins can be used for different industrial and academic purposes such as structure determination, organic synthesis and drug design.

Unit I
Introduction to protein engineering
5 lectures
Protein engineering – definition, applications; Features or characteristics of proteins that can be engineered (definition and methods of study) – affinity and specificity; Spectroscopic properties; Stability to changes in parameters as pH, temperature and amino acid sequence, aggregation propensities, etc. Protein engineering with unnatural amino acids and its applications.

Unit II
Stability of protein structure
5 lectures
Methods of measuring stability of a protein; Spectroscopic methods to study physicochemical properties of proteins: far-UV and near-UV CD; Fluorescence; UV absorbance; ORD; Hydrodynamic properties–viscosity, hydrogen-deuterium exchange; Brief introduction to NMR spectroscopy – emphasis on parameters that can be measured/obtained from NMR and their interpretation.

Unit III
Applications
5 lectures
Forces stabilizing proteins – Van der waals, electrostatic, hydro bonding and weakly polar interactions, hydrophobic effects; Entropy – enthalpy compensation; Experimental methods of protein engineering: directed evolution like gene site saturation mutagenesis; Module shuffling; Guided protein recombination, etc., Optimization and high throughput screening methodologies like GigaMetrix, High throughput microplate screens etc., Application to devices with bacteriorhodopsin as an example; Engineering antibody affinity by yeast surface display; Applications to vaccines, Peptidomimetics and its use in drug discovery.

Recommended Textbooks and References:
5. Journals: (a) Nature, (b) Nature Biotechnology, (c) Applied microbiology and biotechnology, (d) Trends in Biotechnology, (e) Trends in Microbiology, (f) Current opinion in Microbiology, (g) Biotechnology Advances, (h) Genome Research
### Nano-biotechnology

**Credits**

<table>
<thead>
<tr>
<th>Unit</th>
<th>Title</th>
<th>Lectures</th>
</tr>
</thead>
<tbody>
<tr>
<td>Unit I</td>
<td>Introduction to nanobiotechnology</td>
<td>5</td>
</tr>
<tr>
<td>Unit II</td>
<td>Nano – films</td>
<td>5</td>
</tr>
<tr>
<td>Unit III</td>
<td>Nano – particles</td>
<td>5</td>
</tr>
<tr>
<td>Unit IV</td>
<td>Applications of nano – particles</td>
<td>5</td>
</tr>
</tbody>
</table>

**Course Objectives**

The course aims at providing a general and broad introduction to multi-disciplinary field of nanotechnology. It will familiarize students with the combination of the top-down approach of microelectronics and micromechanics with the bottom-up approach of chemistry/biochemistry; a development that is creating new and exciting cross-disciplinary research fields and technologies. The course will also give an insight into complete systems where nanotechnology can be used to improve our everyday life.

**Student Learning Outcomes**

On successful completion of this course, students should be able to describe basic science behind the properties of materials at nanometre scale, and the principles behind advanced experimental and computational techniques for studying nanomaterials.

### Computational approaches

Computational approaches to protein engineering: sequence and 3D structure analysis, Data mining, Ramachandran map, Mechanism of stabilization of proteins from psychrophiles and thermophiles vis-à-vis those from mesophiles; Protein design, Directed evolution for protein engineering and its potential.

### Case studies

Case Studies.

**Recommended Textbooks and References:**

Unit V
Nano – materials
5 lectures
Nanomaterials for catalysis, development and characterization of nanobiocatalysts, application of nanoscaffolds in synthesis, applications of nanobiocatalysis in the production of drugs and drug intermediates.

Unit VI
Nano – toxicity
5 lectures
Introduction to Safety of nanomaterials, Basics of nanotoxicity, Models and assays for Nanotoxicity assessment; Fate of nanomaterials in different stratas of environment; Ecotoxicity models and assays; Life Cycle Assessment, containment.

Recommended Textbooks and References:
1. GeroDecher, Joseph B. Schlenoff, (2003); Multilayer Thin Films: Sequential Assembly of Nanocomposite Materials, Wiley-VCH Verlag GmbH & Co. KGaA
3. Neelina H. Malsch (2005), Biomedical Nanotechnology, CRC Press
5. Recent review papers in the area of Nanomedicine.

Course Objectives
This course will provide students with an overview of current developments in different areas of vaccines.

Student Learning Outcomes
By the end of this course, students should be able to:
• Understand fundamental concepts of human immune system and basic immunology;
• Differentiate and understand immune responses in relation to infection and vaccination;
• Understand requirement and designing of different types of vaccines;
• Understand importance of conventional and new emerging vaccine technologies.

Unit I
Fundamentals of immune system
6 lectures
Overview of Immune system; Human Immune system: Effectors of immune system; Innate & Adaptive Immunity; Activation of the Innate Immunity; Adaptive Immunity; T and B cells in adaptive immunity; Immune response in infection; Correlates of protection.

Unit II
Immune response to infection
9 lectures
Protective immune response in bacterial; viral and parasitic infections; Primary and Secondary immune responses during infection; Antigen presentation and Role of Antigen presenting cells: Dendritic cells in immune response; Innate immune response; Humoral (antibody mediated) responses; Cell mediated responses: role of CD4+ and CD8+ T cells; Memory responses: Memory and effector T and B cells; Generation and Maintenance of memory T and B cells.

Unit III
Immune response to vaccination
8 lectures
Vaccination and immune response; Adjuvants in Vaccination; Modulation of immune responses: Induction of Th1 and Th2 responses by using appropriate adjuvants and antigen delivery systems - Microbial adjuvants, Liposomal and Microparticles as delivery systems; Chemokines and cytokines; Role of soluble mediators in vaccination; Oral immunization and Mucosal Immunity.

Unit IV
Vaccine types & design
3 lectures
History of vaccines, Conventional vaccines; Bacterial vaccines; Viral Vaccines; Vaccines based on routes of administration: parenteral, oral, mucosal; Live attenuated and inactivated vaccine; Subunit Vaccines and Toxoids; Peptide Vaccine.
New Vaccine Technologies; Rationally designed Vaccines; DNA Vaccination; Mucosal vaccination; New approaches for vaccine delivery; Engineering virus vectors for vaccination; Vaccines for targeted delivery (Vaccine Delivery systems); Disease specific vaccine design: Tuberculosis Vaccine; Malaria Vaccine; HIV/AIDS vaccine; New emerging diseases and vaccine needs (Ebola, Zika).

**Recommended Textbooks and References:**

4. Journal Articles (relevant issues) from: Annual Review of Immunology, Annual Review of Microbiology, Current Opinion in Immunology, Nature Immunology, Expert review of vaccines.
<table>
<thead>
<tr>
<th>S.No.</th>
<th>Name of University</th>
<th>Contact Details of Course Coordinator</th>
</tr>
</thead>
</table>
| 1.   | Aligath Muslim University, Aligarh        | Dr. Rizwan Hasan Khan  
Inter-disciplinary Biotechnology Unit  
0571-2720388, 2720165, 09997778669  
Rizwankhan1@yahoo.com Rizwankhan1@gmail.com |
| 2.   | University of Allahabad, Allahabad        | Dr. Shanthy Sundaram  
Centre for Biotechnology  
0532-2545021 (O); 09874614795, 09335158759  
shanthy.cbt@gmail.com |
| 3.   | Banaras Hindu University, Varanasi        | Prof. Arvind Kumar  
School of Biotechnology  
0542-6701584 (Lab) 1590(office) 09473875706(O)  
arvindkumararvind8@gmail.com |
| 4.   | Banasthali Vidyapeeth, Banasthali         | Prof. Vinay Sharma  
Deptt. of Biosciences & Biotechnology  
01438-228302, 228456 (O), 228341/280, 01438 -228365  
vinaysharma30@yahoo.co.uk |
| 5.   | University of Calicut, Calicut            | Dr. P.R. Manish  
Deptt. of Biotechnology  
0494-2407404 (O), 09447760771  
manishramakrishnan123@gmail.com |
| 6.   | Devi Ahilya Viswavidalaya, Indore         | Prof. Anil Kumar  
School of Biotechnology  
0731-240372, 2470373, 2446802 (R)  
ak_sbt@yahoo.com |
| 7.   | Gulbarga University, Gulbarga             | Prof. Ramesh Londonkar  
Deptt. of Biotechnology  
08472-263291,245633, 245337, 09880416391  
londonkarramesh53@rediffmail.com |
| 8.   | Guru Jambheshwar University of Science & Technology, Hisar | Dr. Namita Singh  
Deptt. of Bio and Nanotechnology  
0662-263312, 263165  
namitasingh71@gmail.com |
| 9.   | Guru Nanak Dev University, Amritsar       | Sh. P.K. Verma  
Deptt. of Biotechnology  
0183-2258431, 2258802, 09878366009  
bioetchgndu@yahoo.com |
| 10.  | Himachal Pradesh University, Shimla        | Prof. Reena Gupta  
Deptt. of Biotechnology  
0177-2832154  
reenagupta_2001@yahoo.com |
| 11.  | HNB Garhwal University, Garhwal           | Prof. J. P. Bhatt  
Deptt. of Zoology & Biotechnology  
01370-267322 (O)  
hodzoobiotech@gmail.com; profjpbhatt@gmail.com |
| 12.  | University of Hyderabad, Hyderabad        | Dr. Anand Kondapi  
Deptt. of Biotechnology, School of Life Sciences  
040-23134731, 23134730  
head.dobb@uohyd.ac.in, akksl@uohyd.ernet.in |
| 13.  | Indian Institute of Technology, Mumbai    | Dr. Rinti Banerjee  
Deptt. of Biosciences & Bioengineering  
022-2576 7771, 25767770  
head.bio@iitb.ac.in |
<table>
<thead>
<tr>
<th>S.No.</th>
<th>Name of University</th>
<th>Contact Details of Course Coordinator</th>
</tr>
</thead>
<tbody>
<tr>
<td>14.</td>
<td>Indian Institute of Technology, Roorkee</td>
<td>Dr. Partha Roy</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0332-285686</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:paroylbs@iitr.ac.in">paroylbs@iitr.ac.in</a>, <a href="mailto:partharoy1970@gmail.com">partharoy1970@gmail.com</a></td>
</tr>
<tr>
<td>15.</td>
<td>University of Jammu, Jammu</td>
<td>Dr. Jyoti Vakhlu</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0191-2456534 (O), 09419117624</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:jyotivakhlu@gmail.com">jyotivakhlu@gmail.com</a>, <a href="mailto:jyotivakhlu@jammuuniversity.in">jyotivakhlu@jammuuniversity.in</a></td>
</tr>
<tr>
<td>16.</td>
<td>Jawaharlal Nehru University, New Delhi</td>
<td>Dr. Uttam K. Pati</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Centre for Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>011-26704087, 26704089</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:dean_sbt@mail.jnu.ac.in">dean_sbt@mail.jnu.ac.in</a>, <a href="mailto:uttam@mail.jnu.ac.in">uttam@mail.jnu.ac.in</a>, <a href="mailto:uttamkpati@yahoo.co.in">uttamkpati@yahoo.co.in</a></td>
</tr>
<tr>
<td>17.</td>
<td>University of Kashmir, Srinagar</td>
<td>Prof. Raies Ahmad Qadri</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>09419001315</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:raies.qadri@gmail.com">raies.qadri@gmail.com</a></td>
</tr>
<tr>
<td>18.</td>
<td>Kumaun University, Nainital</td>
<td>Dr. Veena Pande</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>05942-248185</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:veena_kumaun@yahoo.co.in">veena_kumaun@yahoo.co.in</a></td>
</tr>
<tr>
<td>19.</td>
<td>University of Lucknow, Lucknow</td>
<td>Prof. U.N. Dwivedi</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biochemistry</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0522-2740132, 2740148 (O), 09415022445 (M)</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:upendradwivedi@hotmail.com">upendradwivedi@hotmail.com</a></td>
</tr>
<tr>
<td>20.</td>
<td>MS University of Baroda, Baroda</td>
<td>Dr. Mrinalini Nair</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Microbiology and Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0265-2794396/32</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:mnair_in@yahoo.com">mnair_in@yahoo.com</a></td>
</tr>
<tr>
<td>21.</td>
<td>University of Mysore, Mysore</td>
<td>Prof. H.S. Prakash</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Applied Botany &amp; Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0821-2414450, 0821-2411467, 09845488400 (M)</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:hsp@appbot.uni-mysore.ac.in">hsp@appbot.uni-mysore.ac.in</a></td>
</tr>
<tr>
<td>22.</td>
<td>University of North Bengal, Siliguri</td>
<td>Dr. Shilpi Ghosh</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>07602974964, 0353- 2776354</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:ghoshshilpi@gmail.com">ghoshshilpi@gmail.com</a></td>
</tr>
<tr>
<td>23.</td>
<td>North Eastern Hill University, Shillong</td>
<td>Prof. A. Chatterjee</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0364-2722403 (O), 0940213929</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:chatterjeeanupam@hotmail.com">chatterjeeanupam@hotmail.com</a>, <a href="mailto:anupamchatterjee@nehu.ac.in">anupamchatterjee@nehu.ac.in</a></td>
</tr>
<tr>
<td>24.</td>
<td>Pondicherry University, Puducherry</td>
<td>Dr. V. Arul</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology, School of Life Sciences</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0413-2654429, 9444753179</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:head.dbt@pondiuni.edu.in">head.dbt@pondiuni.edu.in</a></td>
</tr>
<tr>
<td>25.</td>
<td>Savitribai Phule Pune University, Pune</td>
<td>Dr. Ameeta RaviKumar</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>020-25691331/4592/1821</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:hdbiotech@unipune.ac.in">hdbiotech@unipune.ac.in</a></td>
</tr>
<tr>
<td>26.</td>
<td>Tezpur University, Tezpur</td>
<td>Dr. S. K. Ray</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deptt. of Molecular Biology &amp; Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>03712-267008/9 (O), 09957184351</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:suven@tezu.ernet.in">suven@tezu.ernet.in</a>, <a href="mailto:suvendra1973@gmail.com">suvendra1973@gmail.com</a></td>
</tr>
<tr>
<td>27.</td>
<td>Utkal University, Bhubaneswar</td>
<td>Prof. Jagneshwar Dandapat</td>
</tr>
<tr>
<td></td>
<td></td>
<td>PG Deptt. of Biotechnology</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0674 (O) 2587389, 0943746087</td>
</tr>
<tr>
<td></td>
<td></td>
<td><a href="mailto:jdandapat.nou@gmail.com">jdandapat.nou@gmail.com</a>, <a href="mailto:j.dandapat@rediffmail.com">j.dandapat@rediffmail.com</a></td>
</tr>
</tbody>
</table>
Core Committee of M.Sc. Biotechnology

Chairperson
1. Dr. Rakesh Bhatnagar, Professor, School of Biotechnology, Jawaharlal Nehru University, New Delhi

Co-chairperson
2. Dr. K. K. Rao, Professor, Department of Biosciences and Bioengineering, Indian Institute of Technology, Bombay

Members
3. Dr. Suman Govil, Advisor, Department of Biotechnology, Ministry of Science and Technology, Government of India
4. Dr. B. J. Rao, Senior Professor, Department of Biological Sciences, Tata Institute of Fundamental Research, Mumbai
5. Dr. Chandrababu, Professor, Centre for Plant Molecular Biology and Biotechnology, Tamil Nadu Agricultural University, Coimbatore
6. Dr. Probodh Borah, Professor, Department of Animal Biotechnology, Assam Agricultural University, Guwahati
7. Dr. Deepak Kaul, Professor, Department of Experimental Medicine and Biotechnology, Postgraduate Institute of Medical Education and Research, Chandigarh
8. Dr. Rajesh Gokhale, Staff Scientist VII, National Institute of Immunology, N. Delhi
9. Dr. Indira Ghosh, Professor, School of Computational and Integrative Sciences Jawaharlal Nehru University, New Delhi
10. Dr. Amitabha Bandyopadhyay, Associate Professor, Department of Biological Sciences and Bioengineering, Indian Institute of Technology, Kanpur
11. Dr. Jitender Verma, CEO, M/s. Lifecare Innovations, New Delhi

Member Secretary
12. Ms. Shreya Malik, Deputy Manager, Biotech Consortium India Limited, New Delhi
Contact Us

DEPARTMENT OF BIOTECHNOLOGY
Block 2, 7th Floor, CGO Complex, Lodhi Road, New Delhi-110003

DR. SUMAN GOVIL
Advisor
suman@dbt.nic.in

BIOTECH CONSORTIUM INDIA LIMITED
5th Floor, Anuvrat Bhawan
210, Deen Dayal Upadhay Marg
New Delhi -110002

DR. PURNIMA SHARMA
Managing Director
ceo.bcil@nic.in

MS. SHREYA MALIK
Deputy Manager
shreya@biotech.co.in

49
Universities/ Institutes offering M.Sc./ M.Tech teaching programmes in biotechnology in India with DBT support

As on May, 2017

M.Sc. GENERAL BIOTECHNOLOGY
M.V.Sc. ANIMAL BIO TECHNOLOGY
M.Sc. MARINE BIO TECHNOLOGY
M.Sc. NEUROSCIENCE
M.TECH. FOOD BIO TECHNOLOGY
M.Sc. ENVIRONMENTAL BIOTECHNOLOGY
M.TECH. PHARMACEUTICAL BIOTECHNOLOGY
M.Sc. BIORESOURCE BIOTECHNOLOGY
M.Sc. AGRICULTURAL BIOTECHNOLOGY
M.Sc. MEDICAL BIOTECHNOLOGY
M.Sc. MOLECULAR & HUMAN GENETICS
M.Sc. INDUSTRIAL BIOTECHNOLOGY
M.TECH. BIO TECHNOLOGY
M.Sc. BIORESOURCE BIO TECHNOLOGY